

## Hemoglobin Electrophoretic Patterns in Punjabi Children With Anemia

Kaur Puneet<sup>1</sup>, Singh Daljit<sup>2</sup>, Kukreja Sahiba<sup>3</sup>, Manjari Mridu<sup>4</sup>

<sup>1</sup>MD – Pathology, Department Pathology

<sup>2</sup>MD – Pathology, Department Pathology

<sup>3</sup>MD – Biochemistry, Department Biochemistry

<sup>4</sup>MD – Pathology, Department Pathology

Sri Guru Ram Das Institute Of Medical Sciences And Research, Amritsar, Punjab

### Abstract

**Context:** Carrier frequency of hemoglobinopathy ranges from 3-17% in Indian population. Early and accurate diagnosis of type of anemia is imperative and requires the use of hemoglobin electrophoresis.

**Aims:** To find the prevalence of hemoglobinopathy and different electrophoretic patterns in Punjabi population in subset of anemic children aged less than 18 years, by using agar gel hemoglobin electrophoresis method.

**Methods and Material:** A prospective study included 100 Punjabi children aged <18 years with hemoglobin <9gm%. Complete hemogram, red blood cell (RBC) indices and peripheral blood film (PBF) was examined. Hemoglobin electrophoresis was performed using agar gel electrophoresis method to find different electrophoretic patterns of hemoglobin in the study group.

**Results:** 100 cases were included with mean age (7.4±5.6years) and mean hemoglobin (7.4±1.4g/dl). Hemoglobin electrophoresis revealed abnormal hemoglobin electrophoretic pattern in five patients. Three patients had an abnormal HbA<sub>2</sub> band (beta thalassemia trait); one patient had abnormal HbF and HbA band (beta thalassemia major); and one patient had an abnormal band in HbSD region (HbS confirmed by sickling). No statistically significant difference was noted in RBC indices, PBF examination in patients with normal versus abnormal hemoglobin pattern.

**Conclusions:** Blood parameters including RBC indices and PBF cannot help diagnose abnormal hemoglobin pattern. Hemoglobin electrophoresis by agar gel electrophoresis method is a cost effective method for screening patients with suspected hemoglobinopathies, however it is time consuming and requires proper standardization.

**Keywords:** Anemia, Hemoglobinopathy, Hemoglobin electrophoresis.

**KeyMessages:** Agar gel electrophoresis is a cost effective method to screen and diagnose hemoglobinopathy in a resource constraint country like India.

### I. Introduction

The burden of hemoglobinopathies in India is tremendous with around 45 million total carriers and approximately 15,000 infants born each year with this gene defect. Carrier frequency of hemoglobinopathy ranges from 3-17% in Indian population. It is imperative to make an early and accurate diagnosis of the type of anemia. Complete blood profile, red blood cell (RBC) indices and peripheral blood film (PBF) do not help to exclude hemoglobinopathy traits and thalassemia from nutritional causes of anemia. Hemoglobin electrophoresis at alkaline pH is the single most important investigation in preliminary screening of abnormal hemoglobin. In a developing country like India, where cost is a major concern, agar gel electrophoresis is an ideal technique for routine screening of abnormal hemoglobins. This study aims to find the prevalence of hemoglobinopathy and different electrophoretic patterns in Punjabi population in subset of anemic children aged less than 18 years, by using agar gel hemoglobin electrophoresis method.

### II. Subjects And Methods

This was a prospective cohort study conducted in the department of Pathology, Sri Guru Ram Das Institute of Medical Sciences & Research, Amritsar. It included 100 anemic children from Punjabi population aged less than 18 years with hemoglobin less than 9 g/dl. Using an automated cell counter, a complete hemogram including RBC indices was obtained and peripheral blood smear was examined in all cases. Hemoglobin electrophoresis was then performed on cases using agar gel electrophoresis as per the following technique:<sup>[4]</sup>

1. 3 ml of whole blood was obtained in an EDTA vials and refrigerated at 4°C.

2. **Hemolysate Preparation:** Red cells washed thrice in excess of normal saline in a test tube. To the washed red cells sediments was added equal amount of distilled water mixed, thoroughly and then equal amount of

solution of carbon tetrachloride was added. The contents were again thoroughly shaken to ensure complete red cell lysis. The lysate thus prepared was then processed.

**3. Buffer Preparation:** A Tris-EDTA-borate (TEB) buffer pH 8.6 was used. It consists of the following

**Stock Buffer**

a. Tris (hydroxy methyl methylamine)	109.00 gms
b. Ethylene Diamine Tetra-acetic acid	1.85 gms
c. Boric acid	30.92 gms
d. Distilled water upto	1 liter

**Working Buffer**

A 1:10 dilution of the stock buffer in distilled water was prepared for routine use.

**4. Agar Gel Preparation:** 500 mg of pure E. Merck agar powder was dissolved in 50 ml of working buffer to give a 1% agar gel. The contents were boiled till they turned uniformly transparent. 5 ml of hot molten agar was poured over an evenly surfaced clean glass slide. The agar was allowed to cool and jellyfy at room temperature.

**5. Charging Of Lysates:** 3-5 µg of hemolysate was collected in thin bore Pasteur pipette and smeared over the edge of a cover slip approximately cut to give a width of 1 cm. The smear edge was then charged briskly into the agar gel both for control and test. The contents were allowed to spread within the slit for a while and the cover slip was withdrawn by a brisk outward movement.

**6.** The charged plates were then placed in an electrophoretic chamber and the electrical circuit was completed by bringing the agar surface into contact with anodal and cathodal chambers of buffer solution through strips of filter paper. A current of 10 mA was passed for 60-90 minutes to give adequate separation of bands.

**7.** The slide thus run was fixed in pure methanol. The preparation was stained with 0.5% amido black solution (in Methanol) for 1-2 minutes. The excess of stain was washed out with 5% acetic acid over a period of 30 minutes. The slide was then dried under hot air current.

**8.** The bands obtained from the patient's hemolysate were compared and matched with the control. Any abnormalities if present were then recorded.

**9.** According to the presence or lack of abnormalities seen in the band; all the patient samples of anemia were segregated as hemoglobinopathic samples and non hemoglobinopathic samples (nutritional or other causes of anemia).

All the cases with abnormal hemoglobinopathic samples were further sub categorized and analyzed.

**III. Results**

Total of 100 cases with male:female ratio of 1.6:1. The mean age distribution of the study population was 7.4±5.6 years with mean age for male 6.3±5.2 years and for female 9.1±5.7 years. Mean hemoglobin in the study group was 7.4±1.4 g/dl. In 85 patients hemoglobin ranged between 6.1 – 8.9 g/dl, fourteen patients had hemoglobin between 3-6 g/dl and only one case was below 3g/dl. Mean haemoglobin value was 7.6±1.3 g/dl for males and 7.2±1.6 g/dl for females. (TABLE 1)

Red blood corpuscular (RBC) count was low in 82 patients in the study group, 31 (81.6%) of female and 51 (82.3%) of male patients had low RBC count. Eight patients had a RBC count more than normal. Low mean corpuscular volume (MCV) was seen in 74 patients, with a mean of 74.0±13.5 fL. A low MCV was seen in 48 (77.4%) of males and 26 (68.4%) females. Normal MCV was seen in 23 patients in the study group while only three patients had a raised MCV. Mean corpuscular hemoglobin (MCH) was less than normal range in 82 cases in the study group with a mean of 23.3±5.3 pg. A low MCH was seen in 49 (79.0%) of males and 33 (86.8%) of females. MCH was normal in 14 patients while four patients had a raised MCH. Mean corpuscular haemoglobin concentration (MCHC) was less than normal in 57 patients in the study group with a mean value of 30.8±2.6 g/dl. A low MCHC was seen in 31 (50%) males and 26 (68.4%) females. 38 patients had a normal MCHC, while it was increased in five patients. Red cell distribution width (RDW) was increased in 94 patients in the study group with a mean RDW of 18.1±3.1%. A raised RDW was seen in 57 (91.9%) males and 37 (97.4%) females. It was normal in the remaining 6 patients. (TABLE 2)

Analysis of PBF revealed a microcytic hypochromic smear in 62 patients, dimorphic smear in 31 patients, normocytic normochromic smear in five patients and macrocytic blood smear in two patients. (TABLE 3)

Amongst the 100 patients in the study group, five patients had an abnormal haemoglobin electrophoresis and 95 had a normal electrophoresis study. Three patients had an abnormal HbA<sub>2</sub> band along with HbA (Beta thalassemia trait) – (Figure 1). One patient had abnormal HbF and HbA band (Beta thalassemia

major) – (Figure 2) and one patient had an abnormal band in the HbSD region – (Figure 3) (HbS confirmed by sickling) – HbS Disease.

#### **Analysis Of Abnormal Electrophoresis Samples**

The mean hemoglobin in patients with beta thalassaemia trait was 8.6g/dl, mean RBC count 5.4million/ $\mu$ l, mean MCV 63.5fL, mean MCH 20.7pg, mean MCHC 32.0g/dl, mean RDW 15.2% with two patients having microcytic hypochromic smear and one patient having dimorphic smear. The patient with beta thalassaemia major on electrophoresis had hemoglobin of 6.1g/dl, RBC count 2.8 million/ $\mu$ l, MCV 72.0fL, MCH 21.3pg, MCHC 29.6g/dl, RDW 26.9% with microcytic hypochromic blood film. The patient with HbS disease had hemoglobin of 8.6g/dl, RBC count 5.5 million/ $\mu$ l, MCV 63.0fL, MCH 21.0pg, MCHC 34.0g/dl, RDW 14.9% with a dimorphic blood film. (TABLE 4)

Overall the mean age of patients with abnormal haemoglobin pattern was 12.2 $\pm$ 4.9 years, mean haemoglobin of 8.0 $\pm$ 1.0g/dl. Of the patients with abnormal haemoglobin electrophoresis 60% were males and 40% were females. The mean RBC count was 4.3 $\pm$ 1.4 million/ $\mu$ l, MCV was 72.4 $\pm$ 14.1fl, MCH was 22.9 $\pm$ 4.2pg, MCHC was 31.4 $\pm$ 1.5g/dl and RDW was 18.1 $\pm$ 4.6%. In the group of patients with abnormal electrophoretic pattern, 60% had microcytic hypochromic smear on peripheral blood film and 40% had dimorphic smear. There was no statistical significant difference of these values in patients with normal versus abnormal hemoglobin electrophoretic pattern. (TABLE 5)

#### **IV. Discussion**

In the 21<sup>st</sup> century, anemia still remains one of the most common cause of morbidity in India, with prevalence of anemia in children in the age group of 6-35 months to be 73.6%.<sup>[5]</sup> Hemoglobinopathy and thalassaemia have a carrier frequency that ranges from 3 to 17% in Indian population. The analysis is based upon 83 population groups from 15 states of India for hemoglobin D, 93 populations for hemoglobin E and 308 populations for sickle cell hemoglobin.<sup>[1]</sup> In a study conducted on sample population of northern India, 12.5% of the study population was found to be having hemoglobin variants with beta thalassaemia trait forming the largest subgroup – to the tune of 8.9%.<sup>[6]</sup>

It is important to make an accurate and early diagnosis of the type of anemia so as to institute appropriate treatment. RBC indices and examination of the peripheral blood film alone does not exclude majority of hemoglobinopathy traits and thalassaemia.<sup>[2]</sup> Hemoglobin electrophoresis at alkaline pH is the single most important investigation in preliminary screening of abnormal hemoglobin. Agar gel electrophoresis is simple, rapid and cost effective method of hemoglobin electrophoresis.<sup>[3]</sup>

Our present study showed the prevalence of hemoglobinopathies in the Punjabi population in children with anemia to be 5%. This is comparable to the study by Balgir who found the prevalence range between 3-17% in Indian population (in 15 states).<sup>[1]</sup> However study by Sachdev et al in Northern India showed the prevalence of hemoglobin variants was 12.5%.<sup>[6]</sup>

The mean values of hemoglobin and RBC indices of cases of beta thalassaemia major in the study by Patel et al in Gujarat is comparable to the present study.<sup>[7]</sup> Shivashankara et al in a study in Karnataka on children with anemia found 15 cases of beta thalassaemia major with mean values of MCV, MCH that is also comparable to the present study.<sup>[8]</sup>

The prevalence of beta thalassaemia trait was 8.9% in study by Sachdev et al.<sup>[6]</sup> The study on 11,000 school children of Delhi and Mumbai done by Madan et al showed that the prevalence of beta thalassaemia trait in Delhi school children was 5.47% as against 2.68% in Mumbai.<sup>[9]</sup> However in the present study the prevalence rate is 3%. We attribute this fact due to the relative small study population in our study. The hemoglobin value in the present study of patients with beta thalassaemia trait was 8.57 g/dl, as against 10.9 – 12.2 g/dl in the study by Madan et al. RBC count, MCV, MCH and MCHC is comparable.<sup>[9]</sup>

In the present study we have one case (1%) with abnormal electrophoretic band in the D, S region which was confirmed to HbS by sickling test. This is low as compared to the average gene frequency of hemoglobin S observed in India (4.3%) as shown by Balgir.<sup>[10]</sup> However Sachdev et al in a study of 2600 cases from North India reported only one case with HbS by HPLC (0.03%).<sup>[6]</sup> Giri et al in a study by agar gel electrophoresis found one out of 51 cases with abnormal hemoglobinopathy to be HbS.<sup>[3]</sup> The mean hemoglobin value, MCHC and RDW of case of sickle cell disease in the present study is comparable to that of study conducted by Patel et al.<sup>[7]</sup>

The mean hemoglobin and RBC indices values of cases of beta thalassaemia trait, beta thalassaemia major and sickle cell disease in the present study are also comparable to the values in similar cases of hemoglobinopathies obtained in a study of electrophoretic patterns conducted by Rao et al in 2010 on 800 suspected hemoglobinopathy cases in AIIMS.<sup>[11]</sup>

In a study done in AIIMS by Tyagi et al where HPLC was performed on all cases to check the accuracy of agar gel electrophoresis, they found good correlation between carefully performed conventional methods and

HPLC—especially in beta homozygous thalassemia, HbE and HbS syndromes. They reported advantage of HPLC over conventional electrophoresis only in limited cases and is a must only in cases with hemoglobin migration in SDG region.

### V. Conclusions

Hemoglobin electrophoresis by agar gel electrophoresis is a cost effective method for screening and diagnosis of patients with suspected hemoglobinopathies as PBF and RBC indices cannot help diagnosis the same; however it is time consuming and requires proper standardization.

### References

- [1]. Balgir RS. The burden of haemoglobinopathies in India and the challenges ahead. *CurrSci* 2000;79:1536-47.
- [2]. Davis L.R. Target cells in hemoglobinopathies. *J ClinPathol* 1972;25:169-70.
- [3]. Giri DD, Patra SB, Patel RZ. Hemoglobin electrophoresis in agar gel - A modified method for routine use. *Indian J PatholMicrobiol* 1984;27:179-83.
- [4]. Tyagi S, Saxena R, Choudhry VP. HPLC- How necessary is it for haemoglobinopathy diagnosis in India? *Indian J PatholMicrobiol* 2003;46:390-3.
- [5]. Technical consultation on “Strategies for Prevention and control of Iron Deficiency Anemia amongst under three children in India”. *Indian Pediatrics* 2002;39:640-7.
- [6]. Sachdev R, Dam AR, Tyagi G. Detection of Hb variants and hemoglobinopathies in Indian population using HPLC: Report of 2600 cases. *Indian J PatholMicrobiol* 2010;53:57-62.
- [7]. Patel J, Patel A, Patel J, Kaur A, Patel V. Prevalence Of Haemoglobinopathies In Gujarat, India: A Cross-Sectional Study. *Int J Hematol* 2009;5:1.
- [8]. Shivashankara A.R, Jaikhani R, Kini A. Hemoglobinopathies in Dharwad, North Karnataka: A Hospital - Based Study. *Journal of Clinical and Diagnostic Research* 2008;2:593-9.
- [9]. Madan N, Sharma S, Sood SK, Colah R, Bhatia HM. Frequency of  $\beta$ -thalassemia trait and other hemoglobinopathies in northern and western India. *India J of Human genetics* 2010;16:16-25.
- [10]. Balgir RS. Genetic epidemiology of the three predominant abnormal hemoglobins in India. *J Assoc Physicians India* 1996;44:25-8.
- [11]. Rao S, Kar R, Gupta SK, Chopra A and Saxena R. Spectrum of haemoglobinopathies diagnosed by cation exchange-HPLC & modulating effects of nutritional deficiency anaemias from north India. *Ind J Med Res* 2010 Nov;132:513-9.

### Legends To Figures

**Figure 1** –Abnormal HbA2 band (Beta Thalassemia Trait)

**Figure 2** – Abnormal HbF and HbA band (Beta Thalassemia Major)

**Figure 3** – Abnormal band in HbSD region (Hb S)

### Tables

**Table 1 - Age & Hemoglobin**

	Mean - Overall	Mean – Males	Mean - Females
Age	7.4 ± 5.6 yrs	6.3 ± 5.2 yrs	9.1 ± 5.7 yrs
Hemoglobin	7.4 ± 1.4 g/dl	7.6 ± 1.3 g/dl	7.2 ± 1.6 g/dl

**Table 2 - Red Blood Cell Indices**

RBC INDICES	Overall	Males	Females
RBC Count	Normal – 10 patients Low - 82 patients	Low – 82.3%	Low – 81.6%
MCV	74.0 ± 13.5 fL	Low – 77.4%	Low – 68.4%
MCH	23.3 ± 5.3 pg	Low – 79.0%	Low – 86.8%
MCHC	30.8 ± 2.6 g/dl	Low – 50%	Low – 68.4%
RDW	18.1 ± 3.1 %	High – 91.9%	High – 97.4%

**Table 3 - Peripheral Blood Film**

PBF	Overall	Males	Females
Microcytic Hypochromic	62 patients	66.1%	55.3%
Dimorphic	31 patients	27.4%	36.8%
Normocytic normochromic	5 patients	6.5%	2.6%
Macrocytic	2 patients	0	5.3%

**Table 4 - Abnormal Electrophoresis Parameters**

Parameters	Beta Thalassemia Trait	Beta Thalassemia Major	Hbs Disease
Numbers	3	1	1
Hb	8.6 G/Dl	6.1 G/Dl	8.8 G/Dl
Rbc Count (Million/ $\mu$ l)	5.4	2.8	5.5
Mcv	63.5fl	72.0fl	63.0fl

Mch	20.7pg	21.3pg	21.0pg
Mchc	32.0 G/Dl	29.6 G/Dl	34.0 G/Dl
Rdw	15.2%	26.9%	14.9%
Smear	Microcytic Hypochromic – 2 Dimorphic - 1	Microcytic Hypochromic – 1	Dimorphic – 1
Abnormal Bands	Hba2	Hbf	Hbs

**Table 5 - Comparison (Normal And Abnormal Electrophoresis)**

Variables	Normal Electrophoresis	Abnormal Electrophoresis
Number	95	5
Age	7.1 Yrs	12.2 Yrs
Hemoglobin	7.4 G/Dl	8.0 G/Dl
Male : Female	1.6:1	1.5:1
Rbc Count	3.2million/ $\mu$ l	4.3million/ $\mu$ l
Mcv	73.9 Fl	72.4 Fl
Mch	23.2 Pg	22.9 Pg
Mchc	30.8 G/Dl	31.3 G.Dl
Rdw	18.1 %	18.1 %
Microcytic Hypochromic	63.2 %	60 %
Normocytic Normochromic	5.3%	0%
Dimorphic	30.5%	40%
Macrocytic	2.1%	0%