

Isotherm Study in Arsenic Tolerant Bacteria Isolated from Arsenic Affected Area in West-Bengal, India

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Abstract: Biosorption of arsenic and iron using arsenic tolerant bacteria i.e. *Bacillus* sp. strain CCBAU 51490 (C-7) and *Bacillus subtilis* subsp. *subtilis* strain DSM 10 (A-2) isolated from halophilic arsenic contaminated soil of Baduria [Arsenic (As) concentration of the soil is 2.18 ppm.] after exposing them to 50 mg/l. and 30 mg/l of arsenic respectively and 42 mg/l. of iron (Fe) in primary and ternary solutions. The Langmuir and Freundlich adsorption isotherm models were used in the equilibrium modeling for determining the ideal condition for arsenic removal from contaminated sludge with this resistant bacterial consortium. Both the adsorption isotherm models were found to fit accurately with the experimental data. The best condition of the biosorption of arsenic by C-7 and A-2 bacteria are 700 µl of the bacterial culture aliquot (3.612×10^8 no. of cells of 0.0607 g. dry wt.) at pH 7 in 50 mg./l arsenic supplemented 30 ml. media at 37°C temperature and 300 µl of the bacterial culture aliquot (1.29×10^7 no. of cells of 0.019 g. dry wt. respectively) at pH 7 in 30 mg./l of arsenic concentration containing 30 ml. media at 37°C temperature, where the bacteria can take up 62.34% and 22.17% of As respectively. This condition satisfies both pseudo-first order and pseudo-second order rate kinetics graphs. The bacterial arsenic biosorption shows the different pattern in presence of iron i.e. C-7 bacteria can uptake 47.08% of As from the ternary solution instead of 62.34% of As from the primary solution and A-2 can uptake 33.7% of As from the ternary solution instead of 22.17% of As from the primary solution after 24 hours.

Keywords: Arsenic and iron biosorption, *Bacillus* sp. strain CCBAU 51490, *Bacillus subtilis* subsp. *subtilis* strain DSM 10, Differential rate kinetics equation, Langmuir and Freundlich adsorption isotherm model.

I. Introduction

Water is the most valuable natural resource in our planet because earth's surface comprises 70% of water and the life on the earth would not exist without it. Although this fact is widely recognized, pollution of water resources is a common problem being faced today (Tarangini, 2009). Heavy metal pollution causes directly by the effluents of industries, refineries and waste treatment plants as well as natural disaster like volcanic eruption followed by low-temperature volatilization. The pollutants enter the water supply from soils/ground water systems, natural water bodies like lakes, rivers and ocean etc. and from the atmosphere via rain water (Vijayaraghavan and Yun, 2008). These pollutants are mainly the toxic and hazardous heavy metals (Vieira and Volesky, 2000), which may accumulate in the organisms under certain environmental condition and cause ecological damage (Jefferies and Firestone, 1984). In case of arsenic EPA (Environmental Protection Agency) has set the arsenic standard for drinking water at 0.010 mg/l (<http://water.epa.gov/drink/contaminants>). Among the different biological methods (for eg. precipitation, adsorption, ion exchange, membrane and electrochemical technologies) bioaccumulation and biosorption have been demonstrated to possess good potential to replace conventional methods for the removal of metals (Volesky and Holan, 1995; Malik, 2004) because it is eco-friendly and cost effective. Both Gram +ve and Gram -ve bacteria can absorb the heavy metals easily due to the metabolically binding of ions onto the cell surface (Binkley & Simpson 2003) and metabolic production of binding protein (Samuelson *et al.* 2000).

The present investigation was undertaken to find the most effective condition of arsenic tolerance of *Bacillus* sp. strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* strain DSM 10 by isotherm modeling to calculate the rate kinetics equation for the biosorption in presence of arsenic and the amount of arsenic uptake was also determined in the same way in the presence of iron.

II. Materials And Methods

2.1. INOCULATION AND BIOSORPTION OF THE BACTERIAL CULTURE

The inoculation and biosorption of the bacterial culture was done according to Pathak and Dikshit, 2011. The bacteria [C-7 and A-2 bacteria Fig. 1(A) and 1(B)] were grown in the nutrient agar medium containing 0.3 gm. of beef extract, 0.5 gm. of peptone, 0.5 gm. of NaCl and 2 gm. of Agar in the 100 ml. of water. Pure

culture were obtained by subculture and pure culturing technique from the isolated colonies. Then they were transferred to the nutrient broth and allowed to grow overnight at 37°C in B.O.D. incubator shaker.

Biosorption studies were done using biomass as a function of various parameters such as

- a) pH
- b) Biomass concentration
- c) Temperature

2.1.1. GROWTH OF THE BACTERIA IN THE PRIMARY SOLUTION

(a) EFFECT OF pH OF THE MEDIA

The bacterial metal sorption was monitored for pH range 5, 7 and 10 (Ferreira and Lund, 1987, Tarangini, 2009). Citrate and bicarbonate-carbonate buffer pH 5 and pH 10 respectively were used for maintaining the proper pH i.e. pH 5 and pH 10 respectively in the media and NaOH and HCl were used as pH regulators. 1/100th part of biomass (v/v) was dispersed in 30 ml of the media containing 50 mg/l and 30 mg/l of arsenic (As) concentration for the two bacterial strains respectively. All flasks were maintained at different pH values i.e 5,7 and 10 for about 24 hours. Solutions were centrifuged (Remi C-24) at 9,000 r.p.m. for 10 minutes and the supernatant was analyzed for the residual concentrations of the metal ions.

(b) EFFECT OF THE BACTERIAL BIOMASS CONCENTRATION

According to Tarangini, 2009, Zhu, 2010 and Babak *et al*, 2012, the effect of the bacterial biomass concentration on the biosorption study was done. Biomass was centrifuged at 9,000 rpm for 10 minutes and different weights of the biomass ranging from 300 µl. to 900 µl. (from 1.548 x 10⁸ to 4.64 x 10⁸ no. of cells in case of C-7 bacteria and 1.29 x 10⁷ to 3.87 x 10⁷ no. of cells in case of A-2 bacteria) aliquots were dispersed in 30 ml. media containing the 50 mg/l and 30 mg/l of arsenic (As) concentration at the optimum pH i.e pH 7 for the two bacterial strains respectively. The solutions were adjusted to the optimum pH in which maximum biosorption of the metal ion occurred by the two bacteria. Flasks were left on the shaker of B.O.D. incubator shaker at 37°C for equilibration. The solutions were later centrifuged at 9,000 r.p.m for 10 minutes (Remi C-24) and the metal ion concentrations were determined from the supernatants using the procedures described later.

(c) EFFECT OF TEMPERATURE OF THE BACTERIAL INCUBATION

According to Tarangini, 2009, Pathak and Dikshit, 2011, Babak *et al*, 2012, the effect of various temperature on the bacterial heavy metal adsorption study was shown. The optimum biomass concentration i.e 700µl. culture aliquot containing 3.612 x 10⁸ no. of cells of 0.0607 g. dry weight in case of C-7 bacteria and 300 µl. culture aliquot containing 1.29x 10⁷ no. of cells of 0.019 g. dry weight in case of A-2 bacteria with optimum pH i.e pH 7 was used to monitor the temperature effect on biosorption of arsenic (As) from the arsenic supplemented media (which was described earlier) by the two bacterial strains. The experiments were carried out at the different temperatures i.e 21°C, 30 °C and 37 °C for C-7 bacteria and 37°C, 30°C for A-2 bacteria and kept the culture flasks the shaker of the B.O.D. incubator shaker, which were set at the different temperatures. The samples were allowed to attain equilibrium. The samples were collected at their stationary growth phase. Then they were centrifuged (Remi C-24) at 9,000 r.p.m. for 10 minutes and the supernatants were analyzed for metal concentration.

(d) ANALYTICAL ESTIMATION OF ARSENIC

The concentrations of Arsenic in the samples were measured by ICP-OES (Thermo Scientific, Model No. ICAP 6000 Duo). The most sensitive lines for arsenic (Sengupta *et al*, 2013) lie in the UV region (193.759 nm.) and an appropriate spectrophotometer would be used.

2.2. ADSORPTION ISOTHERMS

According to Tarangini, 2009 and Zhu, 2010 the optimum biomass of each culture was dispersed in a desired concentration i.e 50mg/l and 30 mg/l of arsenic for C-7 and A-2 bacteria respectively in their optimum medium pH and temperature. The culture flasks were incubated for their respective period of time at the end of which the residual concentrations were determined.

FORMULA AND EQUATIONS USED

The amount of metal bound by the biosorbents i.e the biomass of the bacterial cells was calculated as follows

$$Q = v (C_i - C_f)/m \dots\dots\dots(1)$$

Where Q is the metal uptake (mg metal per g biosorbent), v is the liquid sample volume (ml), C_i is the initial concentration of the metal in the solution (mg/l), C_f is the final (equilibrium) concentration of the metal in the supernatant (mg/l) and m is the amount of the added biosorbent i.e. the dry weight of the bacterial biomass (mg).

The Langmuir model,

$$Q = Q_{\max} \frac{bC_f}{1+bC_f} \dots \dots \dots (2)$$

Where Q_{\max} is the maximum metal uptake under the given conditions, b a constant related to the affinity between the biosorbent and sorbate.

Linearized Langmuir model

$$1/Q = 1/Q_{\max} (1/b C_f + 1) \dots \dots \dots (3)$$

The Freundlich Model,

$$Q = k C_f^{(1/n)} \dots \dots \dots (4)$$

Where k and n are Freundlich constant, which correlated to the maximum adsorption capacity and adsorption intensity, respectively.

Linearized Freundlich equation

$$\log Q = \log k + 1/n \log C_f \dots \dots \dots (5)$$

Equation (1) and (2) can be solved using least square technique, equation (3) using quantile regression, equation (4) using modified least square and equation (5) using computerized method.

2.1.2.(a) GROWTH OF THE BACTERIA IN THE TERNARY SOLUTION

The method was done according to Borrok *et al.*, 2006, Parungao *et al.*, 2007, Tarangini, 2009 and Zhu, 2010. The bacterial seed cultures were inoculated in 30 ml. media supplemented with 50 mg/l and 30 mg/l of arsenic (As) (for C-7 and A-2 respectively) and 42 mg./l of iron (Fe) (the ternary solution) with maintaining the optimum growth condition for the heavy metal uptake. The growth of the bacteria was measured by Colorimetric assay at 580 nm. filter. Then the samples were collected at their stationary growth phase and they were centrifuged (Remi-C24) at 9,000 r.p.m. for 10 minutes. After centrifugation the supernatants were analyzed for the residual As and Fe concentration.

(b) ANALYTICAL ESTIMATION OF ARSENIC AND IRON

The residual concentration of As (as mentioned earlier according to Sengupta *et al.*, 2013) and Fe were measured by ICP-OES (Thermo Scientific, Model No. ICAP 6000 Duo). In ICP-OES, the concentration of iron in the solutions were measured at 259.940 nm. of UV range of the spectrophotometer.

III. Results

3.1(a). EFFECT OF pH OF THE MEDIA

In this study it is observed that the growth of C-7 and A-2 bacteria i.e *Bacillus* sp. strain CCBAU 51490, *Bacillus subtilis* sub sp. *subtilis* strain DSM 10 is much better in pH 7 [Fig. 2(B), 2(F)] medium than pH 5 [Fig. 2(A), 2(E) and pH 10 (Fig. 2(C), 2(G)]. In pH 5 medium because the static phase of the bacteria arrived in just 4 hours and the growth of the bacteria was very low which was measured by the O.D. value in 580 nm. filter. The percentage uptake of As (43.2%, 22.17%) was also higher in pH 7 medium than in pH 10 (35.8%, 8.53%) medium by the such bacteria i.e C-7 and A-2 respectively [Fig. 2(D), 2(H)]. So, pH 7 is the optimum pH for growth and percentage uptake of the metal by the bacteria and it was constant in next experimental parameter i.e biomass of the bacteria.

(b) EFFECT OF THE BACTERIAL CONCENTRATION

The growth of C-7 and A-2 bacteria was higher in in case of dry weight(0.0607g., 3.612×10^8 no. of cells and 0.019 g., 1.29×10^7 no. of cells) of 700 μ l. and 300 μ l aliquot [Fig. 3(C) and 3(F)] than the dry weights of the different aliquots of the both bacteria respectively [Fig. 3(A), 3(B), 3(D) and 3(G), 3(H), 3(I) respectively]. C-7 and A-2 bacteria can uptake 62.34% and 22.17% of As by its 0.0607g. and 0.019 g. of dry weight of 700 μ l. and 300 μ l. aliquot respectively [Fig. 3(E) and 3(J)], which are the highest percentage uptake of As value than the other different biomass of the both bacterial inoculums. So, the optimum condition for the growth and as well as the percentage uptake of arsenic by the both bacteria was found and it remains constant in the next experimental parameter i.e temperature.

(c) EFFECT OF TEMPERATURE OF THE BACTERIAL INCUBATION

The growth and percentage uptake of As by 0.0607g. dry weight of 3.612×10^8 no. of cells of 700 μ l. aliquot of C-7 bacteria and 0.019 g. dry weight of 1.29×10^7 no. of cells 300 μ l. aliquot of A-2 bacteria were the highest in 37°C (62.34% and 22.17% respectively) than in the other different temperatures [Fig. 4(A), 4(B), 4(C) and 4(D) and 4(E), 4(F), 4(G)].

3.2. ADSORPTION ISOTHERM AND RATE KINETICS

C-7 and A-2 bacteria i.e *Bacillus* sp. strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* strain DSM 10 can exhibit an uptake of 62.34% and 22.17% of As respectively. Fig. 6(A), 6(B) depict Freundlich isotherm model of *Bacillus* sp. strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* where the depression zone of the curve is indicated the highest uptake of As in the optimum growth condition of these two bacteria. This condition satisfies both pseudo-first order and pseudo-second order rate kinetics graphs, which are made on the basis of the pseudo-first order and pseudo-second order differential equation $dq_t/dt = k_1(q_e - q_t)$ and $dq_t/dt = k_2(q_e - q_t)^2$ of Yang and Duri, 2005, where q_e (mg g⁻¹) is the solid phase concentration at equilibrium, q_t (mg g⁻¹) is the average solid phase concentration at time t (min), and k_1 (min⁻¹) and k_2 (g mg⁻¹ min⁻¹) are the pseudo-first-order and pseudo-second order rate constants, respectively and from these equations, $\ln(q_e - q_t) = \ln(q_e) - k_1 t$ and $t/q_t = 1/k_2 q_e^2 + t/q_e$ equations are derived, when $t=0$ and $q_t=0$ in the initial condition.

Thus the absorption process of C-7 bacteria and A-2 i.e *Bacillus* sp. strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* strain DSM 10 in its optimum growth condition obeys pseudo-first order rate equation [Fig. 10(A), 8(C)] and as well as pseudo-second order rate equation [Fig. 10(B), 8(D)], while the other growth conditions of the same bacteria obey only pseudo-second order equation (Fig. 7(A), 7(B), 8(A), 8(B), 9(A), 9(B), 11(A), 11(B), 12(A), 12(B), 13(A), 13(B)) because Fig. 10(A) and 8(C) depicts that the logarithm of the difference between uptake of As by C-7 and A-2 bacteria respectively in the equilibrium state (q_e) and the uptake of As by the same bacteria (q_t) at time ' t ' is inversely proportional to the time which means the difference between the uptake of As value by the bacteria is decreased with the time. From Fig. 10(B) and 8(D) it is shown that the uptake of As by the same bacteria (q_t) at time ' t ' is directly proportional to the time ' t ' because q_t increases with increasing time. From the rate kinetics study, the present study indicated that under the optimum growth condition, arsenic adsorption of C-7 reached the equilibrium concentration in 24 hours [Fig. 10(A) and 10(B)] and in this condition arsenic accumulation rate, arsenic uptake and the growth rate of C-7 was highest than the other growth conditions. But in case of A-2, the equilibrium condition was reached in only 2 hours [Fig. 8(C) and 8(D)], at its optimum growth condition which was the most significant than the other conditions.

Fig. 5(A) and Fig. 5(C) depicts the growth of C-7 and A-2 bacteria in presence of both arsenic and iron in its optimum growth condition respectively. From Fig. 5(B) and 5(D) it is suggested that the comparison between the percentage uptake of As and Fe in both primary metal (single metal) solution and ternary metal (mixed metal) solutions which are 62.34% and 22.17% of As and 81.40% and 95.50% of Fe in primary metal solution and 47.08% and 33.7% of As and 78.67% and 43.81% of Fe respectively in ternary metal solutions indicating the stronger competitive sorption of the metals by the bacteria.

IV. Discussion

Adsorption process is usually described through isotherm models, that is, the amount of sorbed particles on the adsorbent materials as a functional effect of its pressure (if gas) or concentration (if liquid) at constant temperature and the quantity adsorbed substances are nearly always accumulated by the mass of the adsorbent to allow comparison of different materials (Foo and Hameed, 2010). According to Ying, C.P., 2007 Foo and Hameed, 2010 adsorption isotherm models are generally in three types; (i) Langmuir model, which suggests as a hypothesis, that is the uptake occurs on a homogeneous surface by monolayer sorption without interaction between sorbed molecules. Langmuir adsorption equation is applicable under the conditions of low pressure. (ii) The Freundlich model proposes a monolayer sorption with a heterogeneous energetic distribution of active sites and with interactions between sorbed molecules, (iii) Redlich and Peterson isotherms present a general isotherm equation in agreement with that of Langmuir and Freundlich isotherm models and (iv) BET Theory put forward by Brunauer, Emmett and Teller explained that multilayer formation is the true picture of physical Adsorption.

Pseudo first order second order kinetic models are the most popular model used to study the biosorption kinetics of heavy metals and to quantify the extent of uptake in biosorption kinetics. Both pseudo first and second order kinetics rate equation determines the decaying of the material at exponential rate. K. Tarangini, 2009 carried the experiment of the biosorption of arsenic, chromium and mercury by the individual and mixed cultures of *Bacillus subtilis* and *Pseudomonas aeruginosa* where 32%, 30% and 28% arsenic, 81.3%, 60.5% and 77.6% chromium, 99.3%, 78.5% and 90.4% mercury were sorbed by the individual and mixed cultures of above mentioned two bacteria from the primary solution (single metal solution). In case of binary solution (two mixed metals solutions) the mixed cultures of two above mentioned bacteria sorbed 30% chromium, 30% mercury and 20.9% arsenic and 70.7% mercury also indicating that the metals microbial interaction and the stronger competitive sorption of the metals by the bacteria. Pathak and Dikshit, 2011 showed the biosorption capabilities of the isolated four bacterial strains on the basis of their fast growth. John and Lara, 2011 showed the increased biosorption of Cd²⁺ from 2.48 to 9.81 mg/g and the maximum monolayer biosorption capacity of *Cassia siamea* was 37.7 mg./g. and they showed the rate kinetics of the Cd²⁺ uptake by the Pseudo-

first order, pseudo-second order and intra particle diffusion model. Parungao *et al.*, 2007 showed the removal of 22% Cu, 24% Cd and 42.75% Pb from the primary solutions (single metal solution) and removal of 16% Cu, 8% Cd and 35% Pb by *Stenotrophomonas maltophilia* (isolated from Mogpog river, Marinduque) from the ternary solutions (mixed metals solution). Borrok *et al.*, 2007 reported that the ternary complexes had the effect of the mobility of aqueous metal cations in natural systems by changing dissolved NOM-metal complexes to colloidal bacteria-metal-NOM complexes. Zhu, 2010 reported that the competitive sorption of Cr and Cu by goethite-*Bacillus thuringiensis*, where Cr had the stronger affinity on the sorbents (Goethite-*Bacillus thuringiensis* than Cu). Babak *et al.*, 2012 showed the affinity order of the metals to the bacteria and they also reported that *Geobacillus thermocatenulatus* had the highest metal sorption capacities than *Geobacillus thermodenitrificans*.

The best effective condition of the biosorption of arsenic by *Bacillus sp.* strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* strain DSM 10 (isolated from arsenic contaminated soil of Baduria, 24 Parganas (North), West-Bengal, India are 0.0607g., 0.019g. dry weight of 700µl, 300µl. of the bacterial culture aliquot at pH 7, 50 mg/l, 30 mg/l of arsenic concentration and 37°C temperature, where the bacterial strain can uptake 62.34% and 22.17% of arsenic respectively (primary solution) which is the highest amount than the other growth conditions of the same bacteria [Fig.6(A), 6(B) Freundlich Isotherm]. So, this condition satisfies both pseudo-first order and pseudo-second order rate kinetics graphs and the absorption process obeys pseudo-first order rate equation and as well as pseudo-second order rate equation, while the other growth conditions of the same bacteria obey only pseudo-second order equation. From this present study it depicts the accumulation rate of As was high in A-2 than C-7 and As uptake capacity was significantly high in case of C-7 due to its cell density. But both bacteria reached their stationary growth phase within 24 hours of incubation. C-7 and A-2 bacteria can also uptake 81.40%, 95.50% of iron from the primary solution in its optimum growth condition and 47.08%, 33.7% of As and 78.67%, 43.81% of Fe respectively from ternary solutions, which indicates the competitive sorption of the two metal by the bacteria

V. Conclusion

The objective of the present study is to find the optimum growth condition, rate kinetics of the heavy metals uptake of the bacteria i.e., *Bacillus sp.* strain CCBAU 51490 and *Bacillus subtilis* subsp. *subtilis* strain DSM 10. These bacterial consortium will be used in the sludge treatment on the basis of their maximum percentage uptake of arsenic and iron from the primary and as well as ternary solution

Acknowledgement

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FIGURES

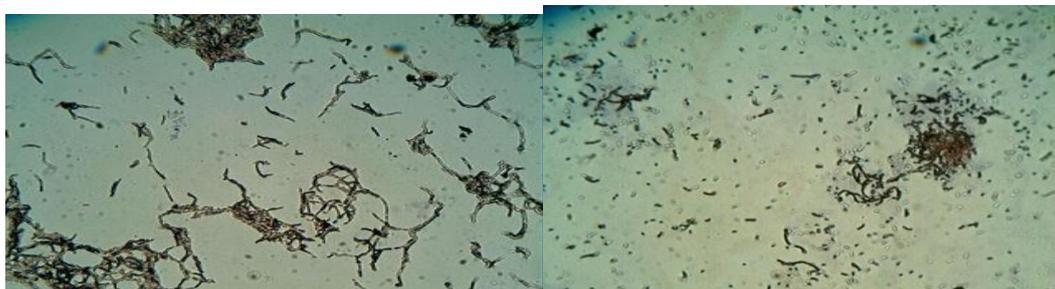


fig. 1(a)

fig. 1(b)

fig.1.(a) and (b). bacterial isolates (C-7) and (A-2) were grown on the nutrient agar media respectively.

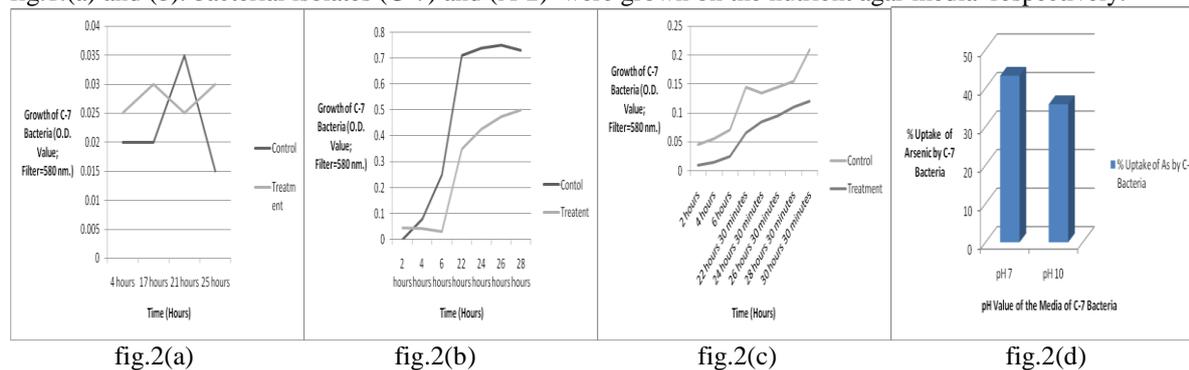


fig.2(a)

fig.2(b)

fig.2(c)

fig.2(d)

fig. 2.(a) growth curve of C-7 bacteria at pH 5 medium in control and treatment condition, (b) growth curve of C-7 bacteria at pH 7 medium in control and treatment condition,(c) growth curve of C-7 bacteria at pH 10 medium in control and treatment condition and (d) % uptake of arsenic by C-7 bacteria at various pH

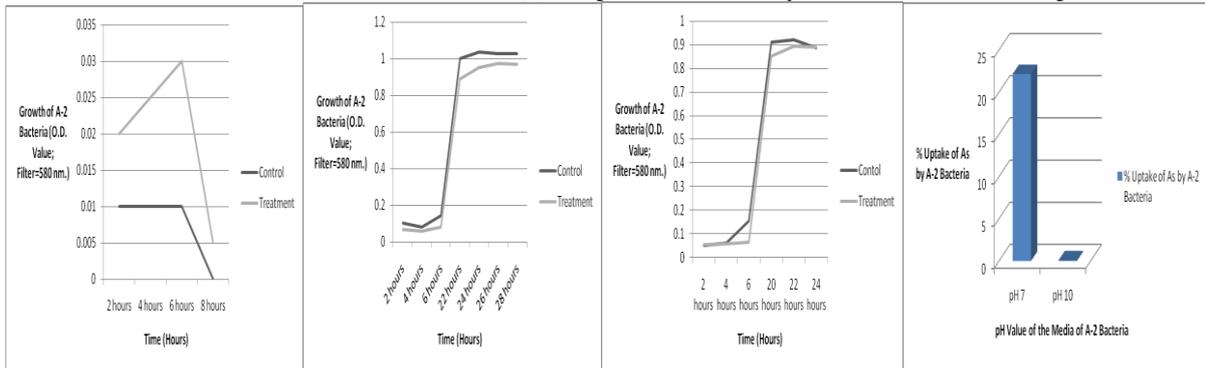


fig.2(e)

fig.2(f)

fig.2(g)

fig.2(h)

fig. 2.(e) growth curve of A-2 bacteria at pH 5 medium in control and treatment condition, (f) growth curve of A-2 bacteria at pH 7 medium in control and treatment condition, (g) growth curve of A-2 bacteria at pH 10 medium in control and treatment condition and (h) % uptake of arsenic by A-2 bacteria at various pH

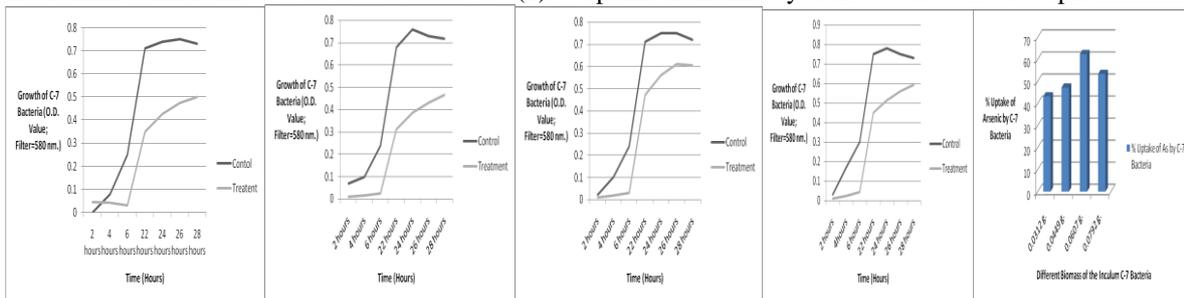


fig. 3(a)

fig. 3(b)

fig. 3(c)

fig. 3(d)

fig.3(e)

fig. 3(a) growth curve of 300µl. aliquot i.e 0.0312g. of C-7 bacteria at pH 7 medium in control and treatment condition, (b) growth curve of 500µl. aliquot i.e 0.0449g. of C-7 bacteria at pH 7 medium in control and treatment condition,(c) growth curve of 700µl. aliquot i.e 0.0607g. of C-7 bacteria at pH 7 medium in control and treatment condition,(d) growth curve of 900µl. aliquot i.e 0.0792g. of C-7 bacteria at pH 7 medium in control and treatment condition and (e) %uptake of arsenic by c-7 bacteria in its different biomass at pH 7 medium

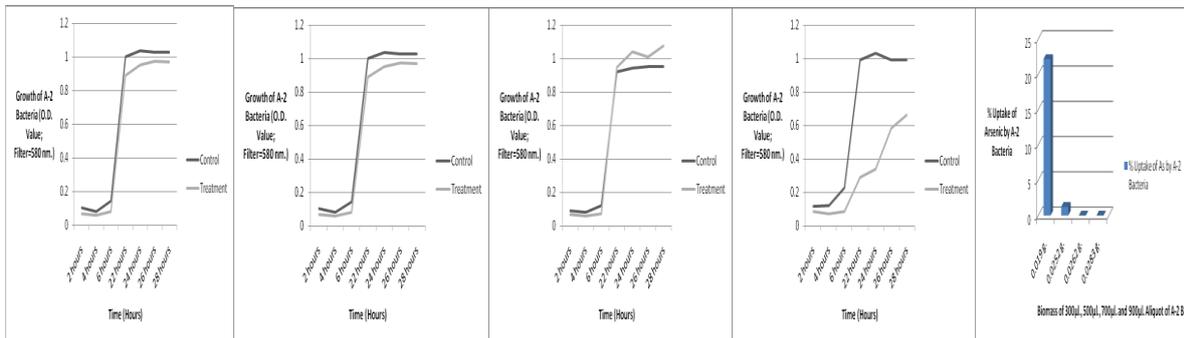


fig.3(f)

fig.3(g)

fig.3(h)

fig.3(i)

fig.3(j)

fig. 3(f) growth curve of 300µl. aliquot i.e 0.0312g. of A-2 bacteria at pH 7 medium in control and treatment condition,(g) growth curve of 500µl. aliquot i.e 0.0449g. of A-2 bacteria at pH 7 medium in control and treatment condition,(h) growth curve of 700µl. aliquot i.e 0.0607g. of A-2 bacteria at pH 7 medium in control and treatment condition,(i) growth curve of 900µl. aliquot i.e 0.0792g. of A-2 bacteria at pH 7 medium in control and treatment condition and (j) %uptake of arsenic by A-2 bacteria in its different biomass at pH 7 medium

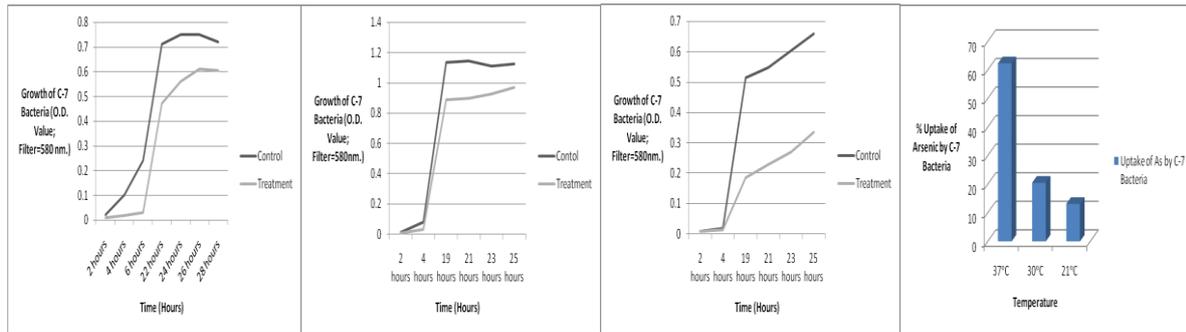


fig. 4(a)

fig. 4(b)

fig. 4(c)

fig. 4(d)

fig. 4(a) growth curve of 700 μ l. aliquot i.e 0.0607g. of C-7 bacteria in pH 7 medium at 37 $^{\circ}$ c in control and treatment condition, (b) growth curve of 700 μ l. aliquot i.e 0.0607g. of C-7 bacteria in pH 7 medium at 30 $^{\circ}$ c in control and treatment condition, (c) growth curve of 700 μ l. aliquot i.e 0.0607g. of C-7 bacteria in pH 7 medium at 21 $^{\circ}$ c in control and treatment condition and (d) %uptake of arsenic by 700 μ l. aliquot i.e 0.0607g. of C-7 bacteria in pH 7 medium at 37 $^{\circ}$ c,30 $^{\circ}$ c and 21 $^{\circ}$ c.

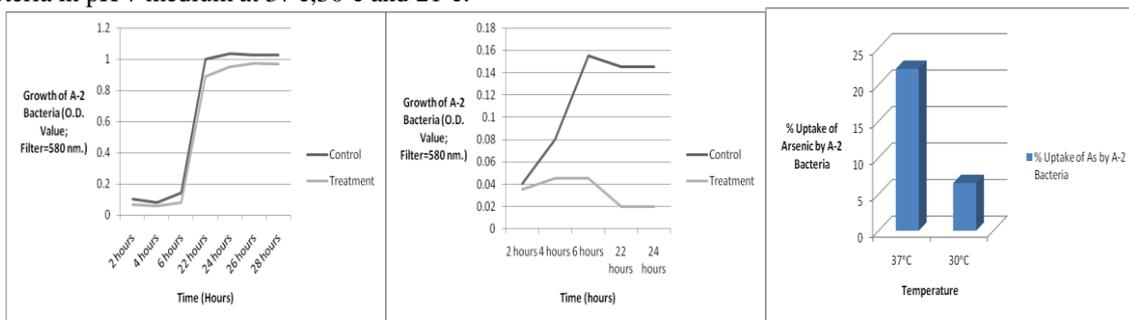


fig.4(e)

fig.4(f)

fig.4(g)

fig. 4(e) growth of 300 μ l. aliquot i.e 0.019g. of A-2 bacteria in pH 7 medium at 37 $^{\circ}$ c in control and treatment condition,(f) growth of 300 μ l. aliquot i.e 0.019g. of A-2 bacteria in pH 7 medium at 30 $^{\circ}$ c and (g) %uptake of arsenic by 300 μ l. aliquot i.e 0.019g. of A-2 bacteria in pH 7 medium at 37 $^{\circ}$ c,30 $^{\circ}$ c.

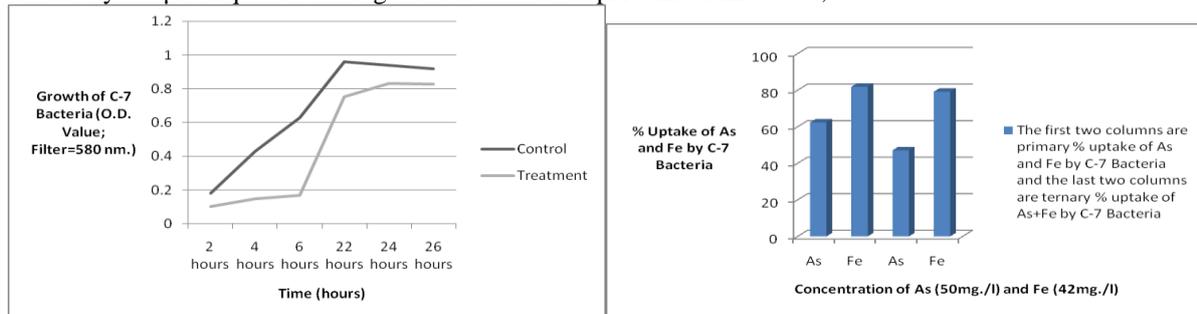


fig. 5(a)

fig. 5(b)

fig. 5(a) growth curve of C-7 bacteria in presence of 50 mg/l of arsenic and 42 mg/l of iron at its optimum pH 7,biomass 0.0607g. of 700 μ l. aliquot and 37 $^{\circ}$ c temperature and (b) %uptake of arsenic and iron by C-7 bacteria from primary solution (single metal solution) and %uptake of arsenic and iron by C-7 bacteria from ternary solution (mixed metals solution).

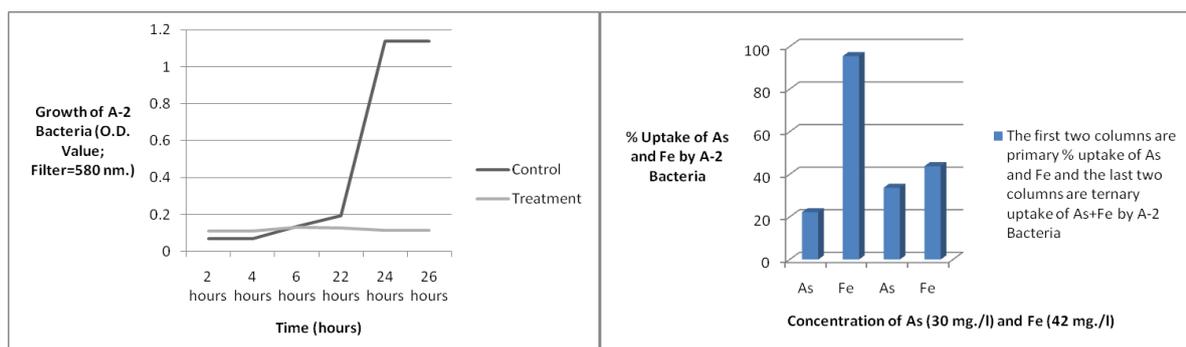


fig. 5(c)

fig. 5(d)

fig. 5(c) growth of A-2 bacteria in presence of 30 mg/l of arsenic and 42 mg/l of iron at its optimum pH 7, biomass 0.019g. of 300 μ l. aliquot and 37 $^{\circ}$ c temperature and (d) %uptake of arsenic and iron by A-2 bacteria from primary solution (single metal solution) and %uptake of arsenic and iron by A-2 bacteria from ternary solution (mixed metals solution).

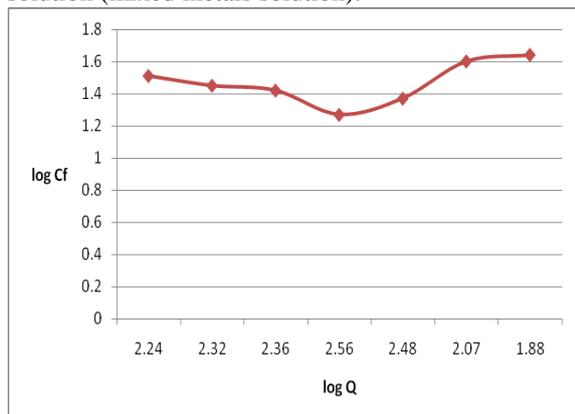


fig. 6(a)

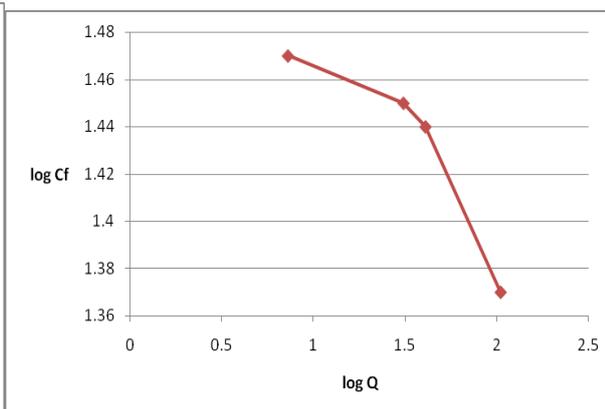


fig. 6(b)

fig.6(a). adsorption isotherm (Freundlich) for arsenic by *Bacillus* sp. strain CCBAU 51490 and 6(b) *Bacillus subtilis* subsp. *subtilis* strain DSM 10

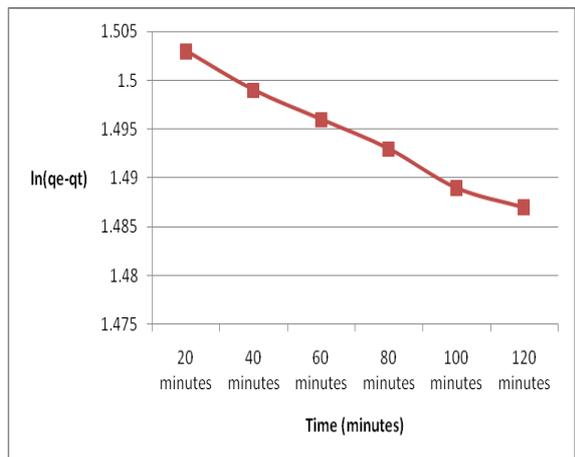


fig. 7(a)

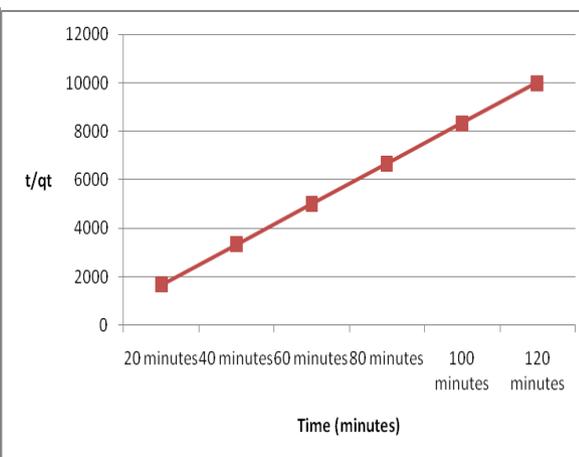


fig. 7(b)

fig. 7(a) first order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.0312g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 10, 37 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.0312g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 10, 37 $^{\circ}$ c temperature.

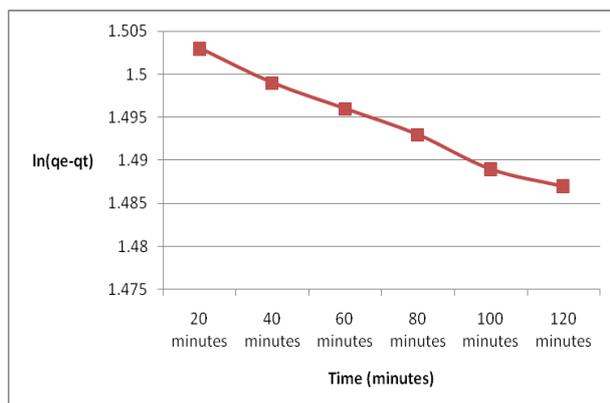


fig. 7(c)

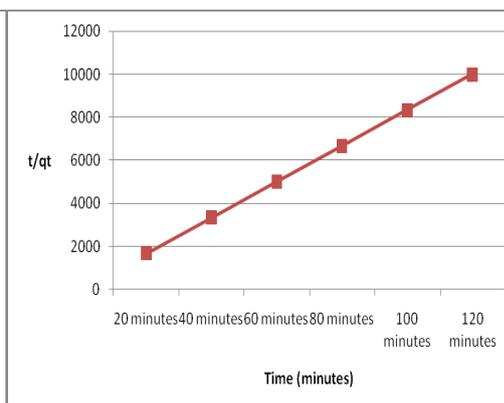


fig. 7(d)

fig. 7(c) first order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.019g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain dsm 10 at 30 mg/l as concentration, pH 10, 37 $^{\circ}$ c temperature and (d) second order kinetics

for arsenic uptake by 300 μ l aliquot i.e 0.019g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain DSM 10 at 30 mg/l as concentration, pH 10, 37 $^{\circ}$ c temperature.

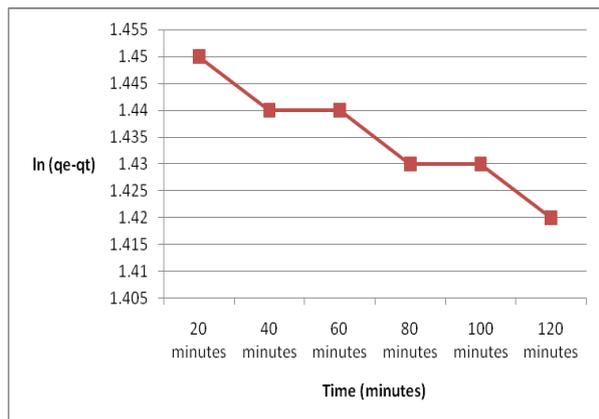


fig.8(a)

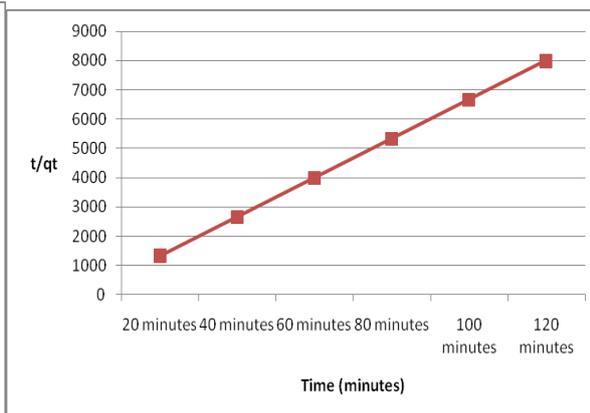


fig. 8(b)

fig. 8(a) first order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.0312g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.0312g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

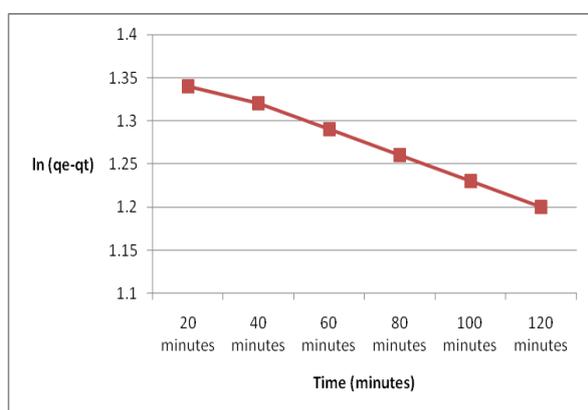


fig. 8(c)

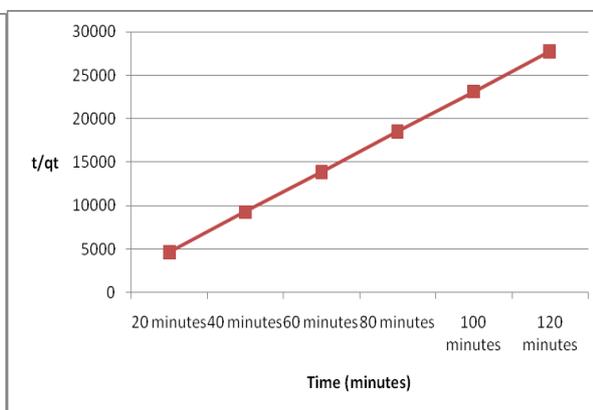


fig. 8(d)

fig. 8(c) first order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.019g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain DSM 10 at 30 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 300 μ l aliquot i.e 0.019g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain DSM 10 at 30 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

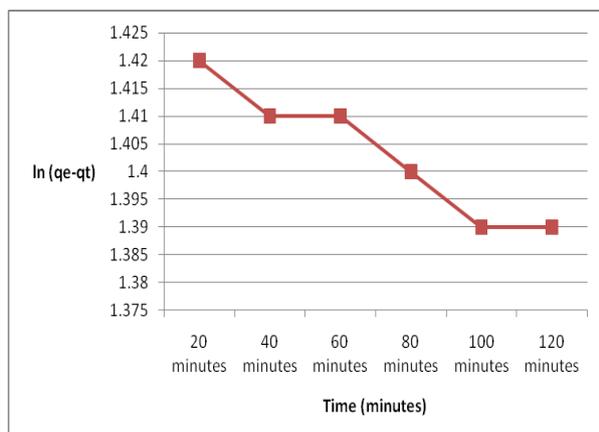


fig. 9(a)

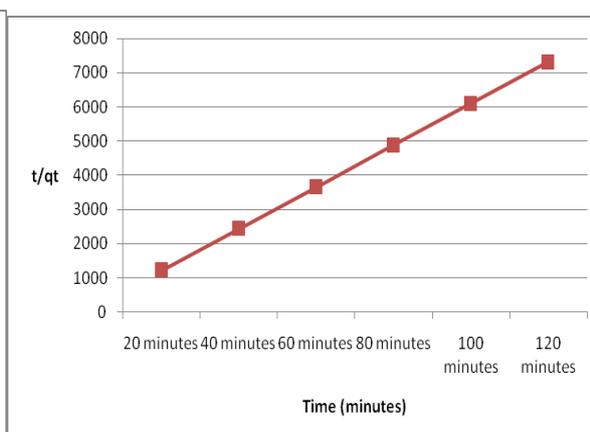


fig. 9(b)

fig. 9(a) first order kinetics for arsenic uptake by 500 μ l aliquot i.e 0.0449g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (b) second order kinetics for

arsenic uptake by 500 μ l aliquot i.e 0.0449g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

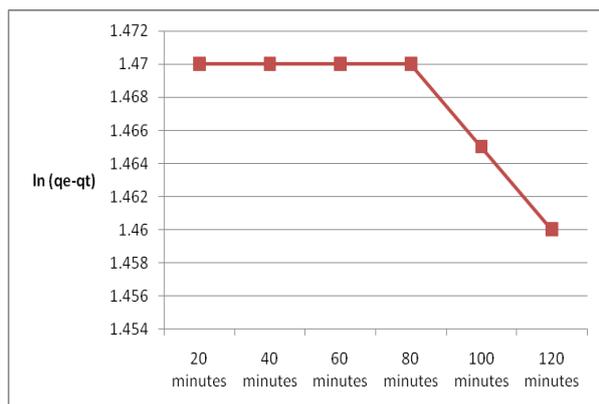


fig. 9(c)

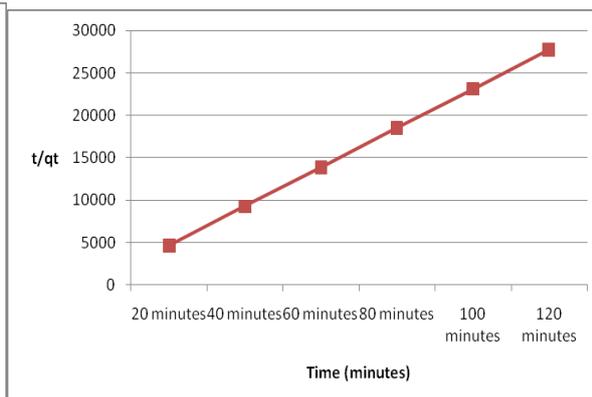


fig. 9(d)

fig. 9(c) first order kinetics for arsenic uptake by 500 μ l aliquot i.e 0.0252g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain DSM 10 at 30 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (d) second order kinetics for arsenic uptake by 500 μ l aliquot i.e 0.0252g. of A-2 bacteria i.e *Bacillus subtilis* subsp. *subtilis* strain DSM 10 at 30 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

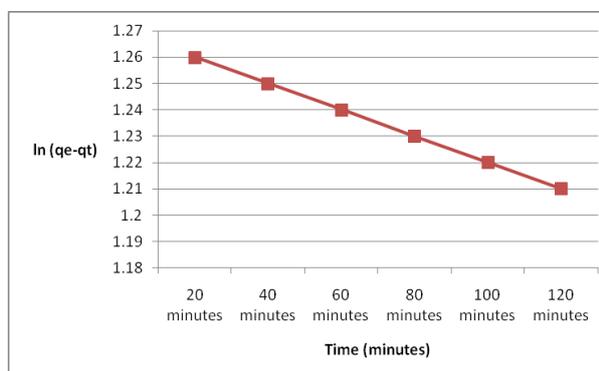


fig. 10(a)

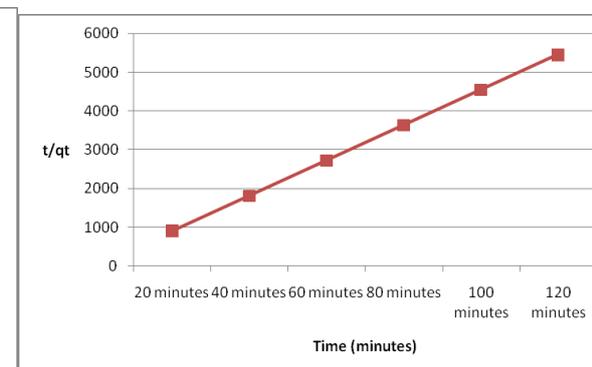


fig. 10(b)

fig. 10(a) first order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of c-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

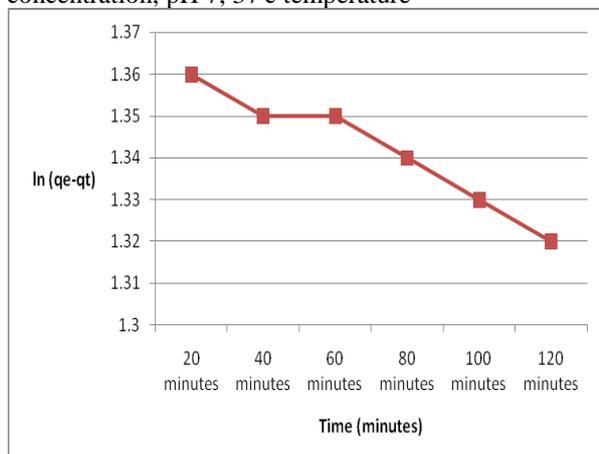


fig. 11(a)

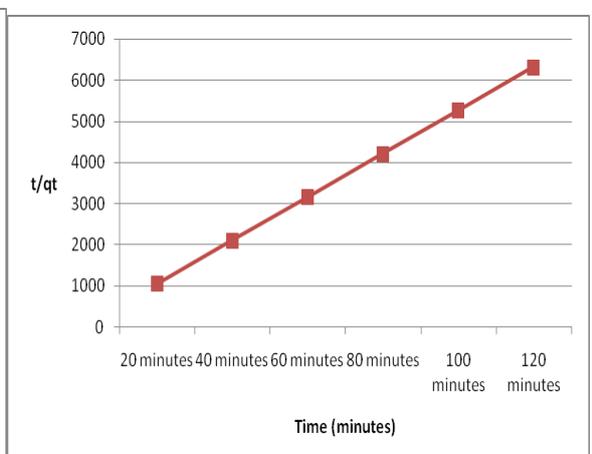


fig. 11(b)

fig. 11(a) first order kinetics for arsenic uptake by 900 μ l aliquot i.e 0.0792g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 900 μ l aliquot i.e 0.0792g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 37 $^{\circ}$ c temperature

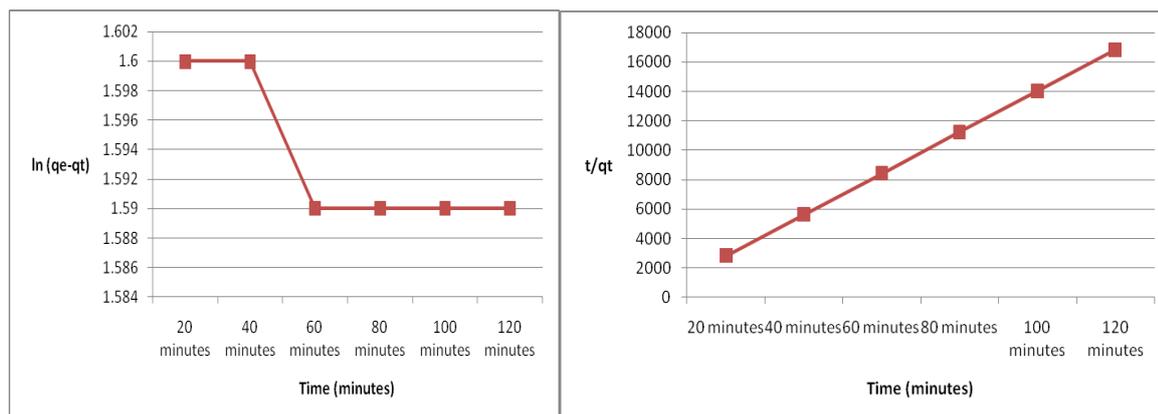


fig. 12(a)

fig. 12(b)

fig. 12(a) first order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 30 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 30 $^{\circ}$ c temperature

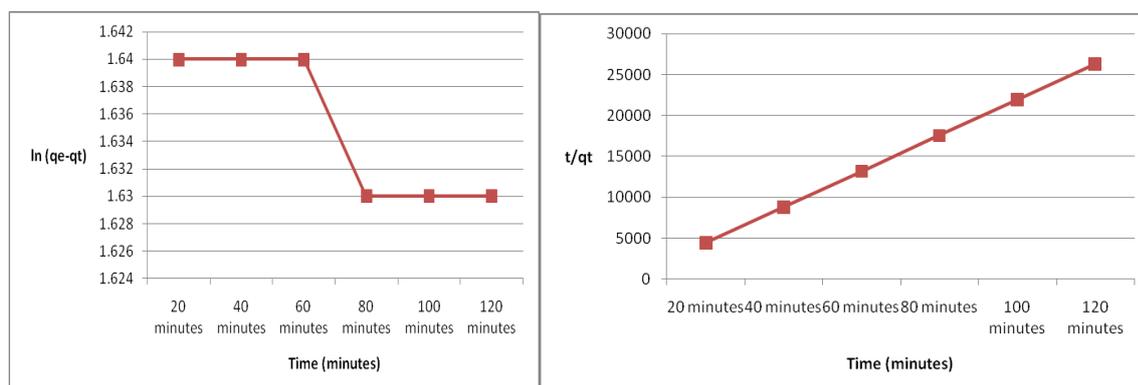


fig. 13(a)

fig. 13(b)

fig. 13(a) first order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 21 $^{\circ}$ c temperature and (b) second order kinetics for arsenic uptake by 700 μ l aliquot i.e 0.0607g. of C-7 bacteria i.e *Bacillus* sp. strain CCBAU 51490 at 50 mg/l as concentration, pH 7, 21 $^{\circ}$ c temperature

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