

Application of chromogen culture media and PCR for detection of *E.coli* O157:H7 and *flicH7*, *rfbO157* Genes respectively in Humans and Dogs isolates

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Abstract: Enterohemorrhagic *E.coli* (EHEC) O157:H7 is a human pathogen responsible for outbreak of bloody diarrhea and hemolytic uremic syndrome worldwide. Polymerase chain reaction was used to investigate the genotypic properties and genetic relationship between *E.coli* O157:H7 obtained from diarrheal humans and dogs cases in Iraq. 88 (88%) *E.coli* isolates were isolated from 100 stool sample of humans and 86 (86%) isolates from 100 fecal samples of dogs by conventional methods (culture, biochemical tests and chrom agar). A total of 174 *E.coli* isolate from human and dog were subjected to PCR to detect the *flicH7* gene and *rfbO157* gene by amplifying a 625-bp region for *flicH7* and a 259 bp region for *rfbO157* gene using the same primers. Results showed that, 4 (4.5 %) and 4 (4.7 %) isolates from humans and dogs respectively give positive product for *flicH7* gene and *rfbO157* gene respectively. These results suggested that, there is a genetic relationship

Keyword: *E. coli* O157:H7, *flicH7* gene, *rfbO157* gene, zoonosis, K9 dogs

I. Introduction

Dogs are the most successful canids, acclimatized to human habitation worldwide. They have contributed to physical, social and emotional well-being of their owners, {10,19}. Domestic dogs have long been recognized to be a potential source of zoonoses for people {21,8}. In particular, zoonotic bacteria and parasites harbored in the canine intestine have been shown to pose a significant risk to human health {21,8}.

People are exposed to these pathogens through direct or indirect contact with infected dogs or their feces, and they may become infected after inadvertent ingestion of a zoonotic agent {16,8}. Although there are more than 300 recognized canine Zoonosis {7,13}, a limited number of these are caused by bacterial agents and have dogs as the main host species, however, these have a significant public health impact as some of them, though to be mild are widespread while others can be severe or even fatal. {6}. such as: *Escherichia coli* (O157:H7).

E. coli O157:H7 and other Enterohemorrhagic serotypes have emerged as major food-borne, zoonotic pathogens in humans, responsible for the hemorrhagic colitis-hemolytic uremic syndrome {17}.

E.coli O157:H7 was the main of serotype EHEC gram-negative, rod (bacillus) belong the family of enterobacteriaceae, and this pathogen was considered a predominant serotype of Shiga toxin producing *E. coli* (STEC) {3,25}.

Lacking studies on *E.coli* O157:H7 recovered from K9 dog especially in Iraq, also for the role of K9 dog in detecting and decreasing terrorist activities, and for many people who are working in management and breeding of K9 dogs, the study aimed to confirm identification of *E.coli* O157:H7 in K9 dogs and humans, and clarify genetic similarity of most important strain which were isolated from the humans and dogs by application of PCR.

II. Methodology

2.1. Collection of samples

One hundred dogs fecal sample, 46 Belgium dogs (5 dogs from Babel and 41 from Baghdad), 3 Black-Wolf dogs from Baghdad, 50 German shepherd-dogs (37 from Baghdad, 12 from salahadeen, and 1 from Babel), 1 Rottweiler dog from Baghdad. Veterinary clinic in Tuz and Ministry of Entry were also sites included in the study.

Also one hundred human's fecal sample which were collected from outpatient of Tuz hospital, suffering from diarrhea. Both humans and dogs fecal samples were collected and transmitted immediately to the laboratory, or by using ice-cooled box to the laboratory for bacterial culture.

2.2. Phenotypic identification by culturing for laboratory diagnosis of *E. coli*

Samples were cultured initially on MacConkey agar and incubated at 37°C for 24 hours, then subcultured on Eosin Methylene blue agar and incubated aerobically at 37 °C for 24 hours {18}.

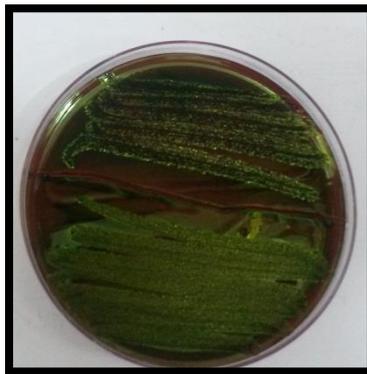


Fig1. *Escherichia coli* on EMB agar

Conventional Biochemical Tests: Biochemical tests were performed according to {5}.

Culturing on CHROM agar

A typical colonies on Sorbitol MacConkey agar (non-Sorbitol fermented *E. coli*) were streaked on chromo agar plates and incubated at 37°C for 24 hours. The colonies of *E. coli* O157 were appeared as mauve colonies while non-pathogenic *E. coli* appeared as blue colonies.

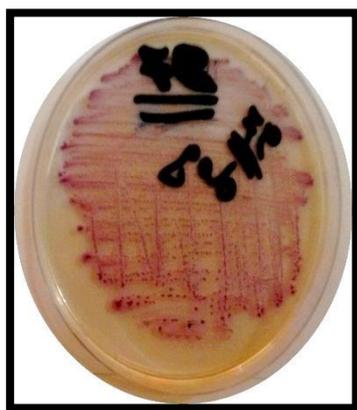


Figure (2): *E. coli* O157:H7 showed Mauve color colonies



Figure (3): *E. coli* showed a blue color On chromo agar O157 at 37°C for 24hrs in chromo agar O157 at 37°C for 24hrs.

2.3 Genotypic identification Polymerase Chain Reaction (PCR) assay:

1: DNA extraction

Genomic DNA of *Escherichia coli* isolate was extracted by using (Genomic DNA Mini Kit, Geneaid. USA)

2: Primers selection

All the primers in this study were obtained from Bioneer, Korea. These primers were used to detect *E. coli* O157:H7 at genus level. These primers were prepared according to the information's of the company. *flicH7* gene was designed by {22}, while *rfbO157* gene was designed by {11}. the sequence of the primers which were used in this study is showed in table(1)

Table(1) sequence of primers for *FlicH7* gene and *rfbO157* gene.

Primer	Sequence(Sequence (5' – 3')		Product Size
<i>flicH7</i>	F	GCG CTG TCG AGT TCT ATC GAG	625 bp
	R	CAA CGG TGA CTT TAT CGC CAT TCC	
<i>rfbO157</i>	F	CGG ACA TCC ATG TGA TAT GG	259bp
	R	TTG CCT ATG TAC AGC TAA TCC	259bp

3. Amplifications:

The PCR amplification mixture (20µl) which was used for detection the genes includes master mix, which provided by Bioneer (Korea) include 5µl of template DNA , 1.5µl of each forwarded and reversed primers and 12µl of nuclease free water to complete the amplification mixture to 20µl.the PCR conditions started with thermo-cycler program, showed in table(2)

Table (2) PCR thermo-cycler condition for *E.coligenes*:

Thermocycling	Primers
	<i>rfbO157</i> <i>flicH7</i>
Initial denaturation	94°C/5min
PCR Amplification cycle*	
Denaturation	94 °C / 1 min.
Annealing	52°C/ 45 sec.
Extention	72 °C/ 90 sec.
Final extention	72°C/10 min.

*Repeat the step (35 cycles)

4:PCR Product Analysis (Agarose Gel Electrophoresis):

It is a very important step to complete PCR assay ,which was used to analyse the PCR product by 1.5%agarose gel electrophoresis supplied with 3µL of ethidium bromide and using 8µL of 100bp ladder, Finally PCR products (bands) were visualized using a UV trans illuminator and photographed by using digital camera.

III. Results and discussion:

1. Phenotyping *E.Coli* O157:H7

The bacteriological culturing revealed a green metallic sheen colonies on eosin methylene blue agar,also these colonies showed mauve color on ChromagarO157 agar, and the bacterial isolates expressed gram-negative stain, this features may be indicated that the bacterial colonies belonged to *E.coli*O157:H7,this result was similar to bacterial colonies of *E.coli*O157:H7 that recorded by {24,1}.

Biochemical tests results were showed as in Table(3),that the isolates were positive for indole test , MR test, oxidase test and catalase test, while they were negative for VP, Simmon’s citrate, and urease tests and acidic yellow in both top and bottom of TSI.

Table (3) shows: - biochemical tests results

Test	Result
indole	Positive
MR	Positive
Catalase	Positive
Simmon Citrate	Negative
VP	Negative
Urease	Negative
TSI	Acidic/Acidic

The bacterial isolation and identification showed that 88(88%), out of 100 human stool samples were *E. coli* positive and 86(86%) out of 100 dog fecal samples *E. coli* positive. After initial isolation and identification of *E. coli* on general, selective and biochemical test.

The isolates of *E. coli* O157:H7 were appeared with mauve color on Chrom agar and other *E.coli* appeared in blue color, Figures (2, 3). Chrom agar aids in diagnosis of *E.coli* O157:H7. *E. coli* O157:H7 utilizes one of chromogenic substrates which produce mauve colored colonies.

While non- *E. coli* O157:H7 organism may utilize chromogenic substrates resulting in blue to blue green colored colonies, this result agreed with the results mentioned by [15,25 and 26]. These isolates were identified and confirmed by multiplex PCR assay, this test showed that 4(4.5%) out of 88 *E. coli* isolates of human stool were *E. coli* O157:H7 positive, while 4(4.7%) out of 86 *E. coli* isolates were *E. coli* O157:H7 positive in dog fecal samples isolates. (14) reported the rate of infection which was higher (3.84 %) among children <2 years of age than other age groups. also (12) recorded 12 (4.2%) persons in the 0-9 age group were infected with this pathogen, in addition (4) mentioned that the highest frequency of diarrheal diseases in the Federal Capital Territory, Abuja occurs within the age group of zero to five years, and this result in agreement with our results as shown in Tables (4,5,6).

Table (4): The prevalence level of diarrheal causative agents in humans stool samples according to gender

Sex	isolated bacteria						% E coli O157:H7
	No	Salmonella enteritidis	% sal	E coli	%E.coli	E coli O157:H7	
female	42	1	2.4	35	83.3	2	4.8
male	58	2	3.4	53	91.4	2	3.4
total	100	3		88		4	

Table (5): The prevalence level of diarrheal causative agents in dog stool samples according to sex

sex	isolated bacteria						% E.coli O157:H7
	No	Salmonella enteritidis	% sal	E.coli	%E.coli	E.coli O157:H7	
female	49	1	2.0	42	85.7	1	2.0
male	51	1	2.0	44	86.3	3	5.9
total	100	2		86		4	

Table(6): The prevalence level of diarrheal causative agents in human stool samples according to Age

Age (year)	isolated bacteria						% E coli O157:H7
	No	Salmonella enteritidis	% sal	E coli	%E.coli	E coli O157:H7	
<1	6	0	0.0	6	100.0	0	0.0
1_4	22	2	9.1	17	77.3	2	9.1
5_14	32	1	3.1	32	100.0	2	6.3
15_45	39	0	0.0	30	76.9	0	0.0
>45	4	0	0.0	4	100.0	0	0.0
total	103	3		89		4	

Genotyping

PCR result of *E. coli* O157:H7

The PCR assay was used to confirm the results of Chrom agar, PCR assay showed that all 8 isolates expressed (*rfbO157*) gene, 4 human stool isolates and 4 dog fecal isolates, also 4 isolates of human stool isolates showed (*flicH7*) gene, and 4 isolates from dog fecal isolates showed (*flicH7*), (Fig: 4).(Fig: 5)

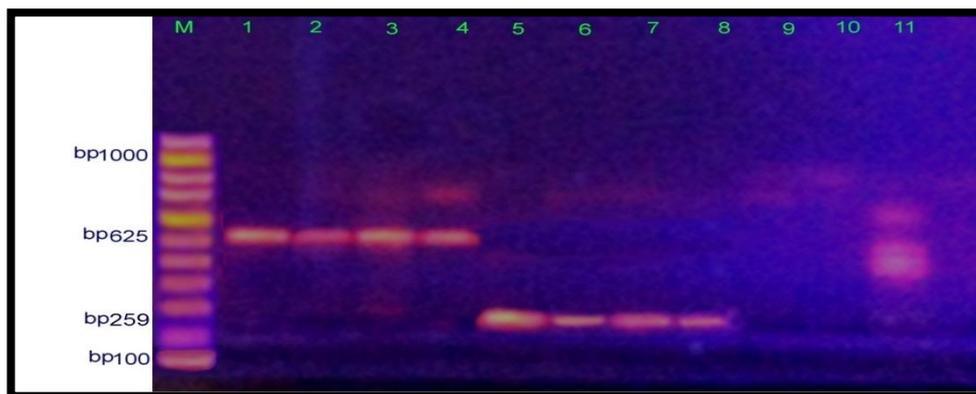


Figure (4): Agarose gel (2%) electrophoresis shows amplification of 259 bp& 625 bp fragments of (*rfbO157*) and (*flicH7*) genes by multiplex PCR. Lanes: 1, 2, 3, 4, 5, 6, 7, and 8 positive amplification of *E. coli* O157:H7 for human. Lane M: 100 bp DNA marker.

Four isolates of *E. coli* O157:H7 from each origin *E. coli* out of (88) human stool isolates and (86) dog stool isolates of *E. coli*. these results agreed with {23}, who reported that PCR assays are proven specific and sensitive in detecting microbial pathogens such as *E. coli* O157:H7. Also {20} mentioned that gene based method such as PCR technique is more reliable than biochemical and serological tests for diagnosis of *E. coli* O157:H7. The main advantage of the employed PCR method is its ability to detect rough isolates or the isolates having a masked O antigen {9}.

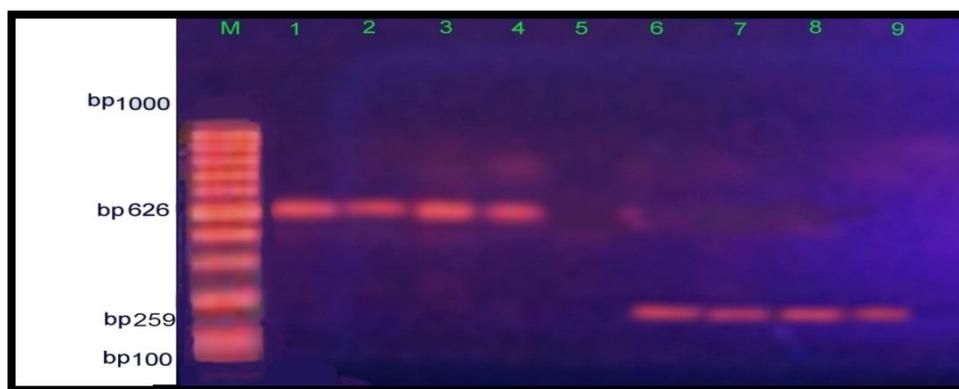


Figure (5): Agarose gel (2%) electrophoresis shows amplification of 259 bp& 626 bp fragments of (*rfbO157*) and (*flicH7*) genes by multiplex PCR. Lanes: 1,2,3,4,6,7 and 8 positive amplification of *E. coli* O157:H7 for dog feces. Lane M: 100bp DNA marker

IV. Conclusion

The study concluded that there is a genetic relationship between human and dog's isolates, which was confirmed by conventional biochemical tests and PCR amplification targeting the *flicH7*, *rfbO157* of *E. coli* O157:H7.

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