

Chemical composition and antimicrobial activity of the essential oil of wild *Thymus vulgaris* grown in South Jordan

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Abstract: There is a high variability in chemical composition of essential oil from *Thymus vulgaris* grown in different countries and different localities in the same country. Only three reports were published on the essential oil composition of *Thymus vulgaris* grown in Jordan and non of them reported from South Jordan. The current study aims to determine the essential oil composition of *Thymus vulgaris* grown wild in South Jordan and to test their activity against clinical isolate antibiotics resistant bacteria. The composition of the essential oil from *Thymus vulgaris* was determined using GC-MS while the screening for essential oil activity was carried out using disc diffusion method. The results of GC-MS produced forty-eight components of the oil representing more than 97% of the oil contents, monoterpenes being most abundant (about 85%) with thymol (37.05%), *cis*-dihydrocarvone (9.34%), carvacrol (8.45%), hydroxy-3-(3-methyl-2-butenyl)-3-cyclopenten-1-one (8.41%), *p*-cymene (5.73%), *cis*-sabinene (4.42%), *z*-isoeugenol (3.342%), and aromadendrene (3.42%) as major constituents.

The essential oil from *Thymus vulgaris* was found to be active against all the tested bacteria except in *Pseudomonas aeruginosa*. The high concentrations of thymol and carvacrol may account for its high activity against resistant bacteria.

Keywords: GC-MS, resistant bacteria, thyme, thymol, carvacrol.

I. Introduction

Essential oils are composed of volatile aromatic compounds with strong odor produced by plants as secondary metabolites. They are usually obtained by steam distillation [1]. The use of essential oils in medicines, perfumes, cosmetics and food preservatives is as old as civilization [2]. Different classes of compounds that may be present in essential oils include 1- Terpenes which include: monoterpene and sesquiterpenes, 2-Oxygenated compounds including phenols, alcohols, aldehydes and ketones, esters.

The chemistry of essential oils is influenced by the local geography and weather conditions [3], as well as the season and time of day when the plants are harvested and how they are processed, packaged and stored. A plant such as *Thymus vulgaris* L (Lamiaceae) (thyme) grown in one area might produce an essential oil with a different chemistry than thyme grown in another location. Thyme is an aromatic plant commonly used in the Mediterranean countries including Jordan as a herbal tea, flavoring agent and spice and a medicinal plant [4]. Extracts from thyme have been used in traditional medicine for the treatment of several respiratory diseases like asthma and bronchitis [5]. *Thymus* species have been shown to have strong antibacterial, antifungal, antiviral, antiparasitic, spasmolytic and antioxidant activities [4].

In recent decades, the incidence of hospital-acquired infections with antibiotic-resistant bacteria has increased remarkably. Notable amongst these infections is methicillin-resistant staphylococcus aureus (MRSA) [6, 7]. Antibiotic resistance organisms are an evolving problem requiring new strategies to combat infection due to these strains. A growing trend is "return to nature" [8]. There is mounting evidence in support of the use of plant-derived essential oils against pathogenic microorganisms [9, 10]. Both clinical and in vitro studies have demonstrated the potent bactericidal properties of some essential oils including efficacy against antibiotic-resistant strains such as MRSA [11].

The aim of the present work was to determine the essential oil composition of *Thymus vulgaris* L. type grown wild in Jordan and to test their activity against clinical isolate antibiotics resistant bacteria.

II. Materials And Methods

2.1 Collection and authentication of plants

Fresh amount of the *Thymus vulgaris* was collected from Mutah, South Jordan, during flowering period and vegetative phase. The plant materials were taxonomically identified and authenticated by the Botanical Survey of Yarmouk University.

2.2 Isolation of essential oil

Aerial parts (100 g) of *Thymus vulgaris* were finely chopped and subjected to hydrodistillation for 4 h using a Clevenger-type apparatus, yielding 1.2%, pale yellowish oil. Subsequently, oil was dried over anhydrous sodium sulfate and immediately stored in GC-grade hexane at 4°C until the analysis by gas chromatography/mass spectrometry (GC/MS) were carried out.

2.3 Essential oil composition

GC–FID analysis: The oils were analyzed in an Agilent (Palo Alto, USA) 6890N gas chromatograph fitted with a 5% phenyl–95% methylsilicone (HP5, 30 m × 0.25 mm × 0.25 μm) fused silica capillary column. The oven temperature was programmed to elevate from 60°C to 240°C at 3°C/min, and hydrogen was used as carrier gas (1.4 mL/min); 1.0 μL of a 1% solution of the oils in hexane was injected, in split mode (1:30). The injector was kept at 250°C and the flame ionization detector (FID) was kept at 280°C. Concentrations (% contents) of oil ingredients for *Thymus vulgaris* were determined using their relative area percentages obtained from GC chromatogram, assuming a unity response by all components.

GC–MS analysis: Chemical analysis of the essential oils was carried out using gas chromatography–mass spectrometry (Agilent (Palo Alto, USA) 6890N gas chromatograph). The chromatographic conditions were as follows: column oven program, 60°C (1 min, isothermal) to 246°C (3 min, isothermal) at 3°C/min, the injector and detector temperatures were 250°C and 300°C, respectively. Helium was the carrier gas (flow rate 0.90 mL/min) and the ionization voltage was maintained at 70 eV. A HP-5 MS capillary column (30 m × 0.25 mm i.d., 0.25 μm film thicknesses) was used. A hydrocarbon mixture of alkanes (C₈–C₂₀) was analyzed separately by GC-MS under same chromatographic conditions using the same HP-5 column. Kovats Retention Indexes (KRIs) were calculated by injection of a series of n-alkanes (C₈–C₂₀) in the same column and conditions as above for gas chromatography analyses.

Identification of the oil components were based on computer search using the library of mass spectral data and by comparison of their calculated Kovats retention index (KRI) with those of the available authentic standards and literature data.

2.4 Maintenance and preparation of cultures

Eight clinical isolates antibiotics resistant bacteria were used in this study. Four strains of Gram positive bacteria: Methicillin-resistant *Staphylococcus aureus* (MRSA), Methicillin-sensitive *Staphylococcus aureus* (MSSA), *Staphylococcus epidermidis*, *Streptococcus pyogenes*, and four strains of Gram negative bacteria: *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Pseudomonas aeruginosa*, were studied. Isolates were purified on specific nutrient agar plates and characterized by the use of standard microbiological and biochemical methods like Gram stain, catalase test, coagulase test and an API system (bioMérieux, France).

The bacteria were incubated at 37°C for 24 h by inoculation into broth. Inoculums (1 mL) per plate containing 10⁶ bacterial cell/ml were spread on Mueller Hinton agar (Oxoid, Hampshire, England).

2.5 Disc diffusion assay

The antibacterial activity of the thyme essential oil was determined by the disc diffusion method according to the National Committee for Clinical Laboratory Standards. Sterile paper discs of 6 mm in diameter were impregnated with 5 μL essential oil and deposited on the agar surface. Petri dishes were placed at 4°C for 2 h to facilitate the dissemination of extract on the culture medium, and then incubated at 37°C for 24 h. Negative water control and positive antibiotic disc control were used for each sample. At the end of the period, inhibition zones formed on the medium are evaluated in mm. Studies were performed in triplicate.

III. Results and Discussion

3.1 Chemical composition of the essential oil

The freshly isolated *Thymus* essential oil was a yellow liquid with yield 1.2%. The identified components of the essential oils, their percentages and retention indices are listed in Table 1.

The components of essential oil were divided into five classes: hydrocarbon monoterpene, oxygenated monoterpenes, hydrocarbon sesquiterpene, oxygenated sesquiterpenes and diterpene (Table I).

Based on GC and GC–MS analysis of the essential oil of thyme, 48 components were identified representing 97.59% of the total detected constituents. The major constituents of the oil were thymol (37.05%), cis-dihydrocarvone (9.34%), carvacrol (8.45%), hydroxy-3-(3-methyl-2-butenyl)-3-cyclopenten-1-one (8.41%), p-cymene (5.73%), cis-sabinene (4.42%), z-isoeugenol (3.342%), aromadendrene (3.42%) as major constituents. The results shows that the concentrations of thymol and carvacrol is high and higher than reported by other researchers [12-15].

Table 1: Constituents (%) of the essential oil of *Thymus vulgaris*

KRI	Compound	%A
864	2E-Hexenal	0.55
873	E-2-Hexen-1-ol	0.12
904	2,5-Dimethyltetrahydrofuran	0.12
931	α -Thujene	0.77
940	α -Pinene	0.52
958	Camphene	0.18
981	Sabinene	0.11
990	trans-Isolimonene	0.36
999	delta-2-Carene	1.51
1011	3-Octanol	0.52
1018	p-Menth-1(7),8-diene	0.24
1021	α -Terpinene	1.72
1026	p-Cymene	5.73
1028	Limonene	0.73
1031	1,8-Cineole	0.41
1037	beta-(Z)-Ocimene	0.10
1072	cis-Sabinene	4.42
1073	para-Mentha-3,8-diene	0.14
1085	para-Mentha-2,4(8)-diene	0.10
1097	Terpinolene	0.16
1105	2,5-Dimethylstyrene	0.10
1111	β -Linalool	0.10
1118	β -Thujone	0.25
1190	cis-Dihydrocarvone	9.34
1258	Carvenone	0.96
1271	cis-Carvone oxide	1.54
1295	Thymol	37.05
1304	Carvacrol	8.45
1352	Eugenol	0.20
1379	2-Ethyl-5-propylphenol	0.30
1380	Carvacrol acetate	0.10
1399	z-Isoeugenol	3.42
1402	2-Methyl-5-(1-methylethyl)- acetate Phenol	0.14
1412	Hydroxy-3-(3-methyl-2-butenyl)-3-Cyclopenten-1-one	8.41
1422	β -Caryophyllene	0.10
1436	Cinerolon	0.65
1444	Aromadendrene	3.42
1476	γ -Murolene	0.32
1510	γ -Cadinene	0.30
1520	7-epi- α - Selinene	2.21
1580	Spathulenol	0.50
1590	Caryophyllene oxide	0.59
1592	Globulol	0.13
1610	5-epi-7-epi- α -Eudesmol	0.10
1631	β -Cedrene epoxide	0.11
1654	Cedr-8(15)-en-9-alpha-ol	0.12
1668	Valerianol	0.14
1963	Isophyllocladene	1.16
	Hydrocarbon monoterpenes	12.23
	Oxygenated monoterpenes	72.74
	Hydrocarbon sesquiterpenes	6.35
	Oxygenated sesquiterpenes	2.33
	diterpene	1.16
	Miscellaneous	2.78
	Total	97.59

3.2 Antimicrobial activity

The results in Table 2 show that Gram-positive bacteria were more sensitive to *Thymus vulgaris* essential oil than Gram negative bacteria. All tested Gram positive bacteria were sensitive while in Gram negative bacteria *E.coli* and *Proteus mirabilis* were sensitive while *Klebsiella pneumoniae* showed a moderate sensitivity and *pseudomonas aeruginosa* was resistant. The high content of *Thymus vulgaris* essential oil with thymol and carvacrol may account for its high antibacterial activity.

Table 2. Antibacterial activity of thyme essential oil

Name of Bacteria used	Zone of inhibition of Thyme essential oil in mm	Antibiotic used	Zone of inhibition in mm
MRSA	24	Vancomycin	19mm
MSSA	24	Vancomycin	18mm
S.epid	26	Vancomycin	21mm

Strep. pyog	26	Clindamycin	26mm
E.Coli	26	Chloramphenicol	28mm
Klebsla. pneu	12	Chloramphenicol	23mm
Pseudo. aeru	2	Ceftazidime	25mm
Proteus mirabilis	23	Ampicillin	14mm

Each test was assayed in triplicate and the values for zone of growth inhibition are presented as means of triplicate

The fight against bacterial resistance continues to be a major health concerns worldwide. Specifically MRSA has become a serious public health concern. It attracted the attention of the medical research community for urgent need to develop new drugs to treat bacterial infections. Thyme essential oil may provide a solution to overcome the resistant showed by MRSA.

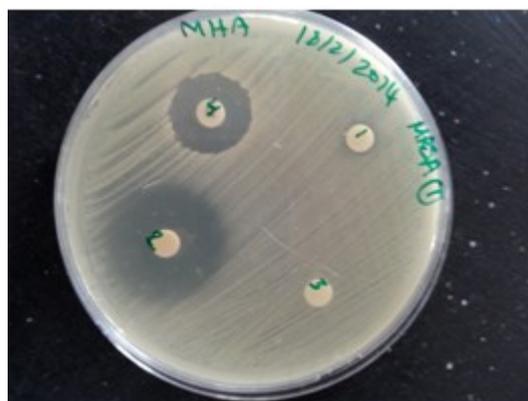


Figure 1: Diameters of inhibition zones (mm) of Thymus vulgaris essential oil against MRSA. (1. Water, 2. Thymus essential oil, 3. Water, 4. Vancomycin)

Thymus essential oil was found to have antimicrobial activity against all microorganisms tested except in *Pseudomonas aeruginosa* which is a known drug resistant bacterium. The same result also was found by Lambert [16]. As to the question: why Thymus essential oil was found to be active against MRSA but not against *Pseudomonas aeruginosa*, the answer may lie in its composition. Thymus essential oil contains thymol and carvacrol which are considered to be its most active ingredients against bacteria although the rest of the component may also be active [17-22]. Thymol and carvacrol are known to disrupt the cell membrane causing its structural and functional damage [16]. *Pseudomonas aeruginosa* has a highly impermeable outer membrane whereas MRSA does not. Impermeability of outer membrane makes it more difficult for thymol and carvacrol to enter inside the bacterial cell. In addition, while *P. aeruginosa* has a highly active efflux pump that can very quickly pump foreign compounds back out of the cell before they can cause any damage; MRSA does not [23].

IV. Conclusion

This study paper showed that the essential oil of *Thymus vulgaris* was active against antibiotic resistant bacteria including MRSA so that essential oils may represent a cheap, safe and effective treatment option against antibiotic-resistant pathogens.

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